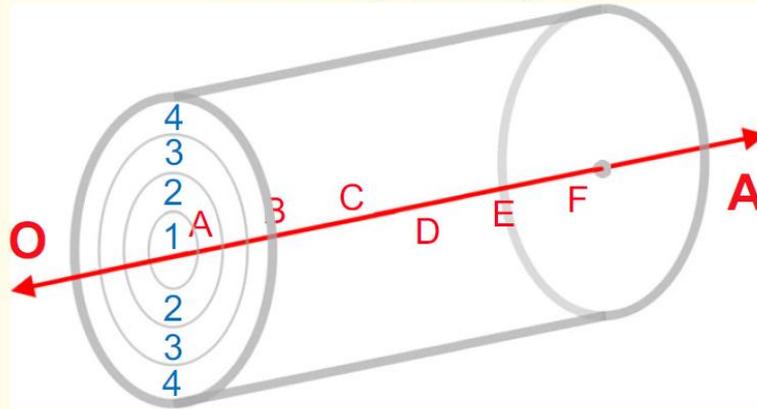


Fig. 1. The distribution of different body symmetries among animals. Alternative scenarios that can explain the emergence of bilaterality are depicted. An-Veg, animal-vegetal; DS-VS, dorsal surface-ventral surface; O-A, oral-aboral; A-P, anterior-posterior; D-V, dorsal-ventral.

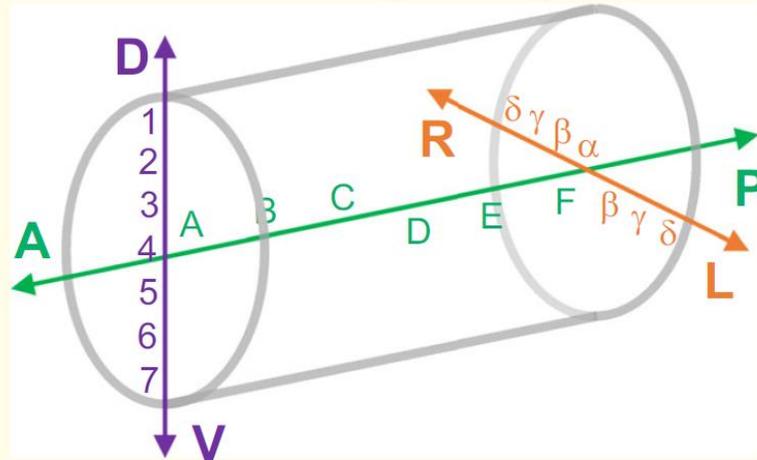
Box 2. Bilaterality and the diversification of body plans

Radial symmetry



$$6 \times 4 = 24 \text{ unique coordinates}$$

Bilateral symmetry



$$6 \times 7 \times 4 = 168 \text{ unique coordinates}$$

Eje Antero-posterior

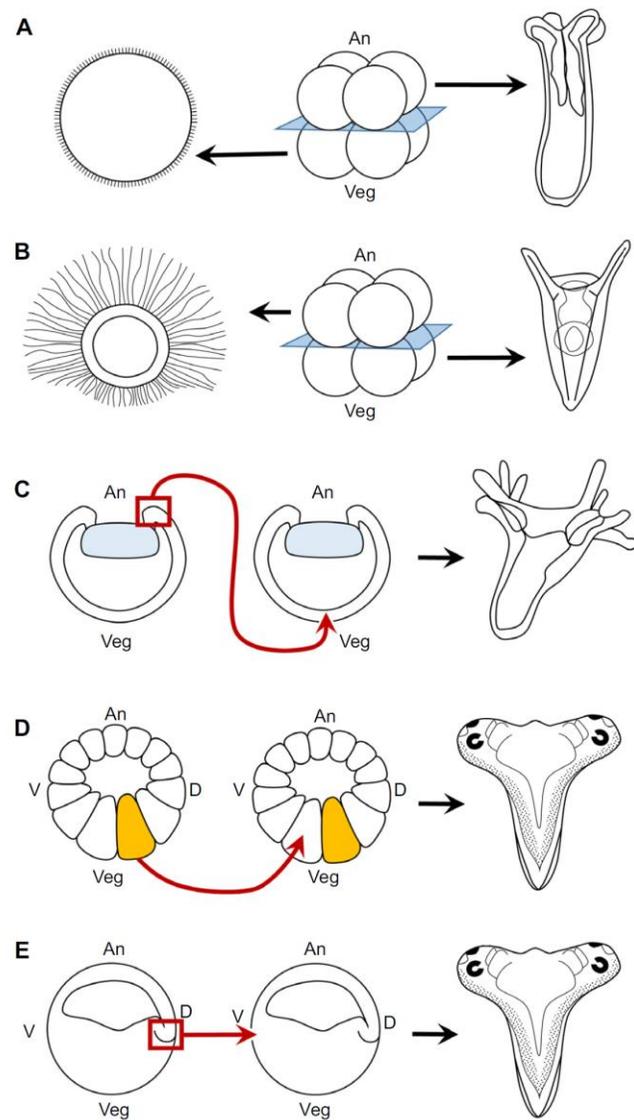
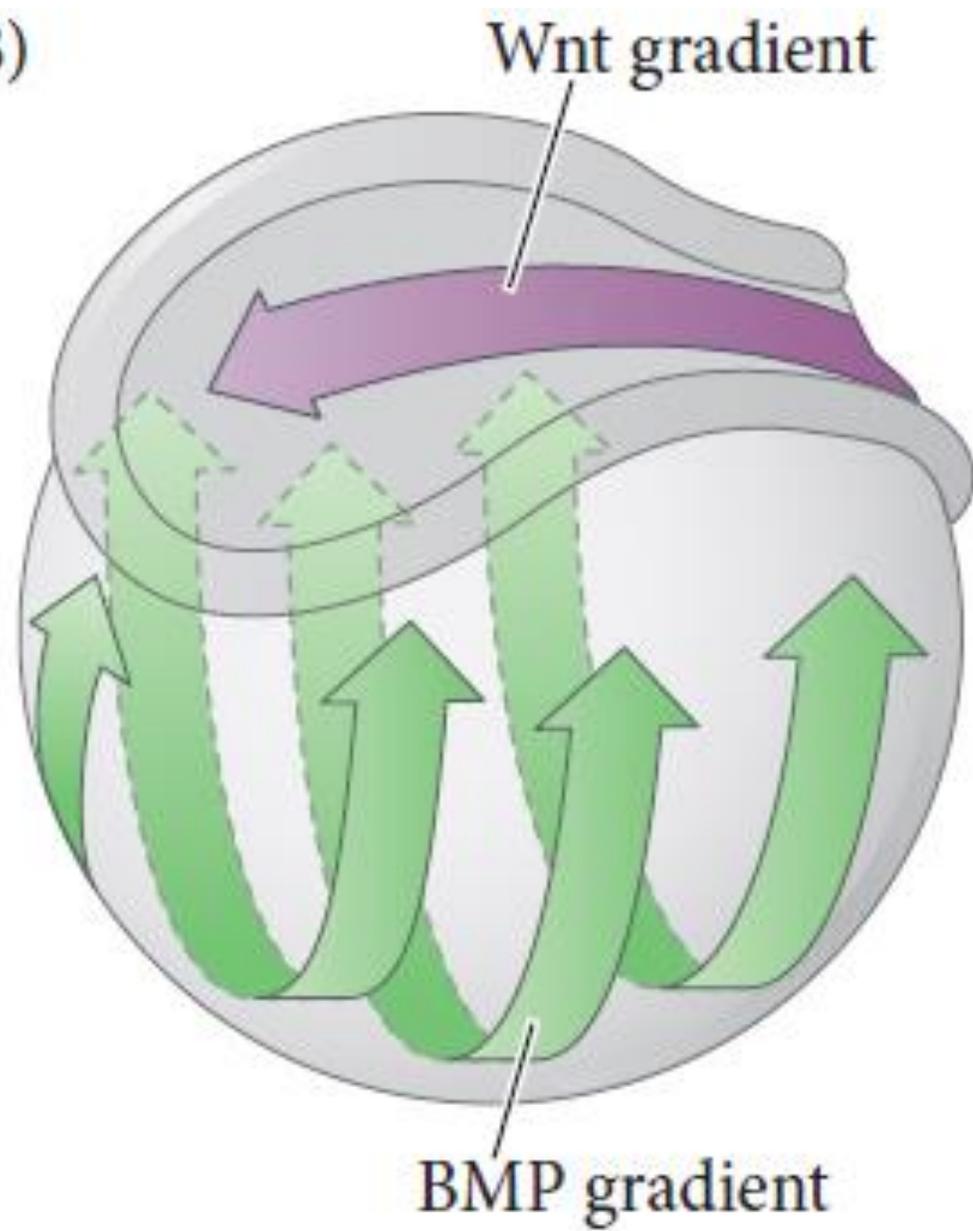


Fig. 3. β -Catenin signalling-dependent axial organizer capacity.
 (A,B) Bisection experiments in *Nematostella* (A) and sea urchin (B) demonstrate the presence of axial determinants in the cytoplasm inherited by cells giving rise to the gastrulation pole. For both *Nematostella* and the sea urchin, a primary polyp (A) or a pluteus larva (B) with normal body axes develops from only the half of the embryo where gastrulation would normally take place. (C,D) Transplantations of the blastopore lip of *Nematostella*, just like transplantations of the amphibian Nieuwkoop centre (D) or Spemann-Mangold organizer (E) into a ventral position, result in the formation of the Siamese twin embryos. An, animal pole; Veg, vegetal pole; D, dorsal; V, ventral.

(B)



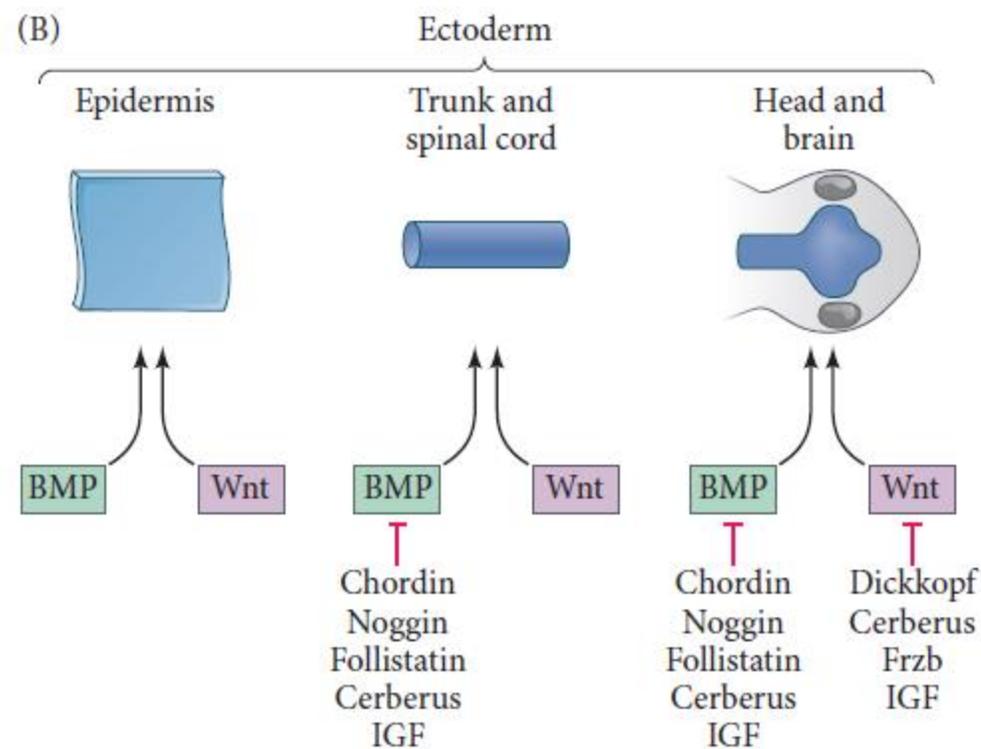
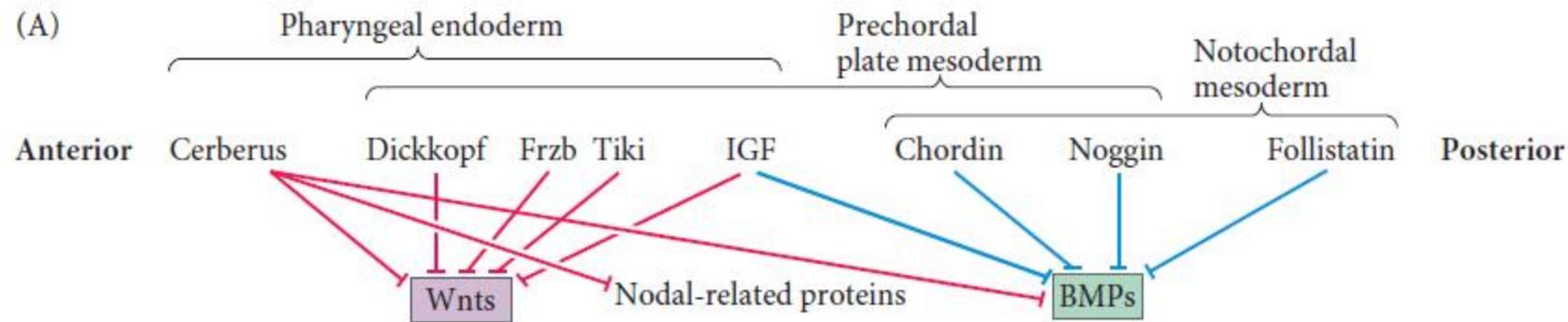
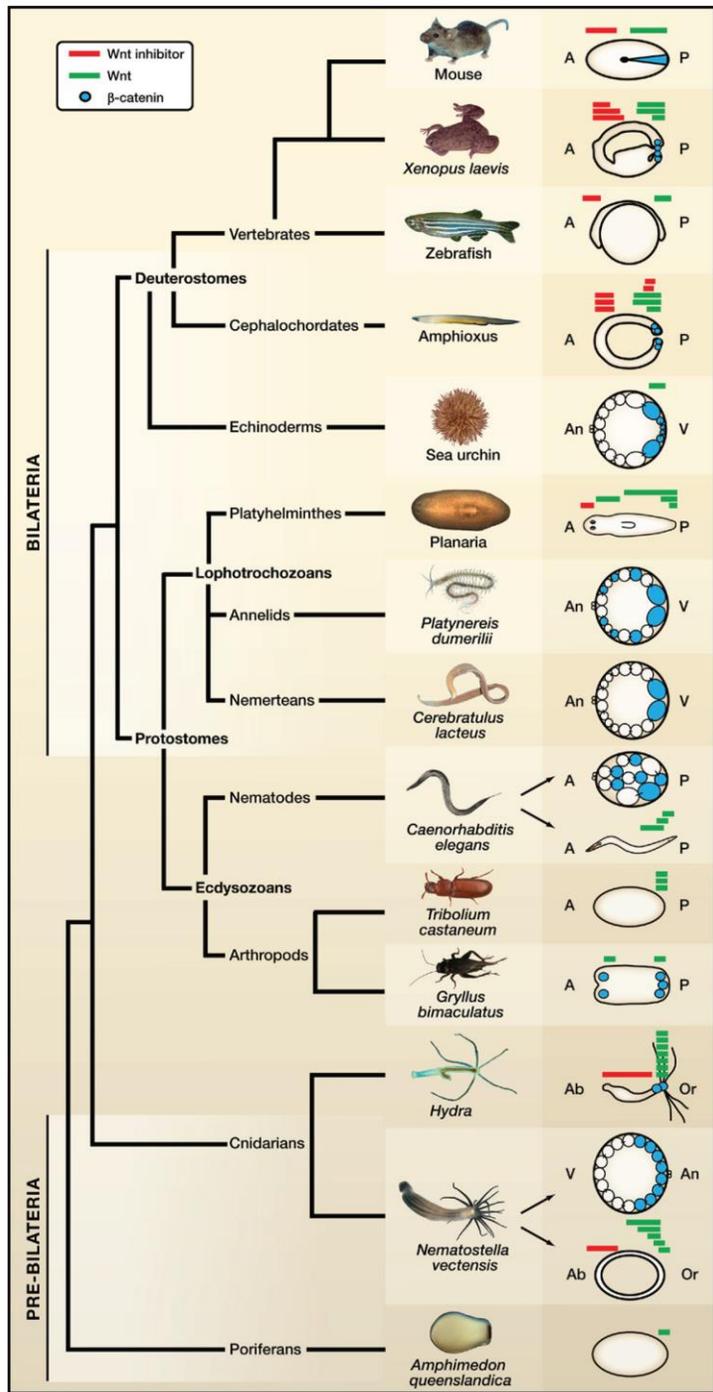
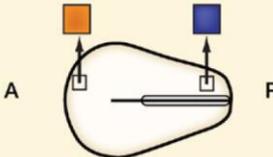
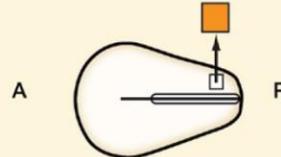
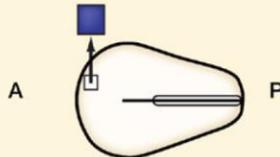
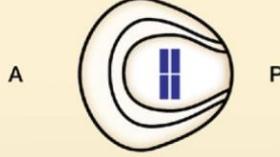
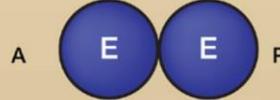
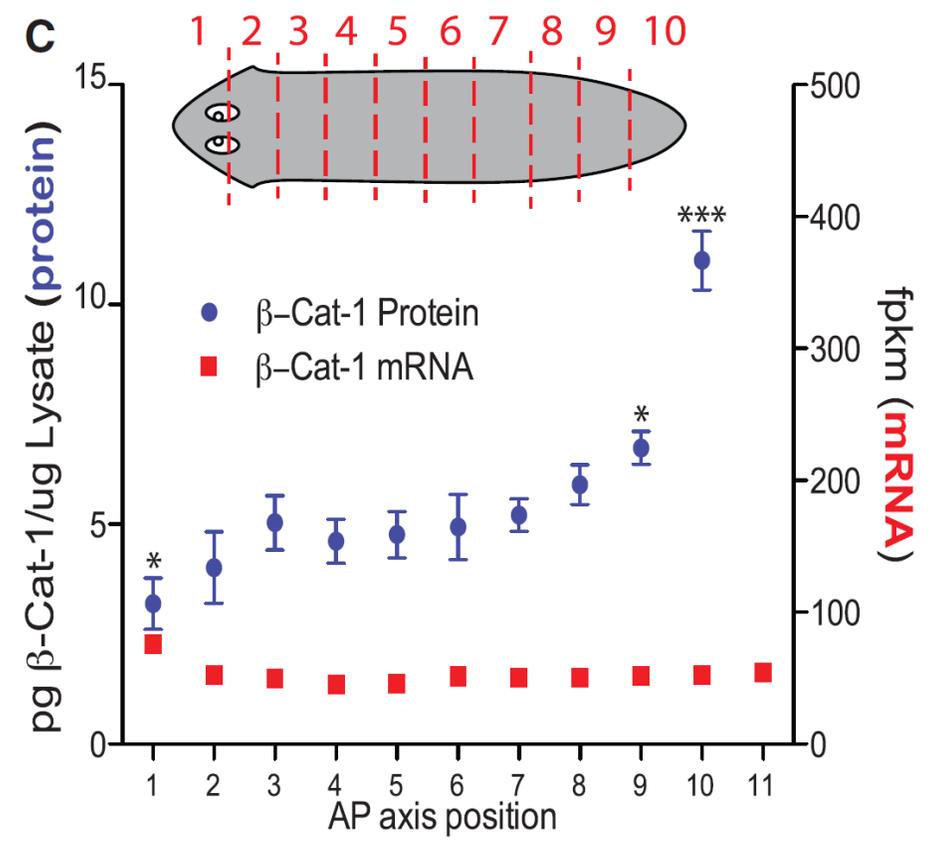
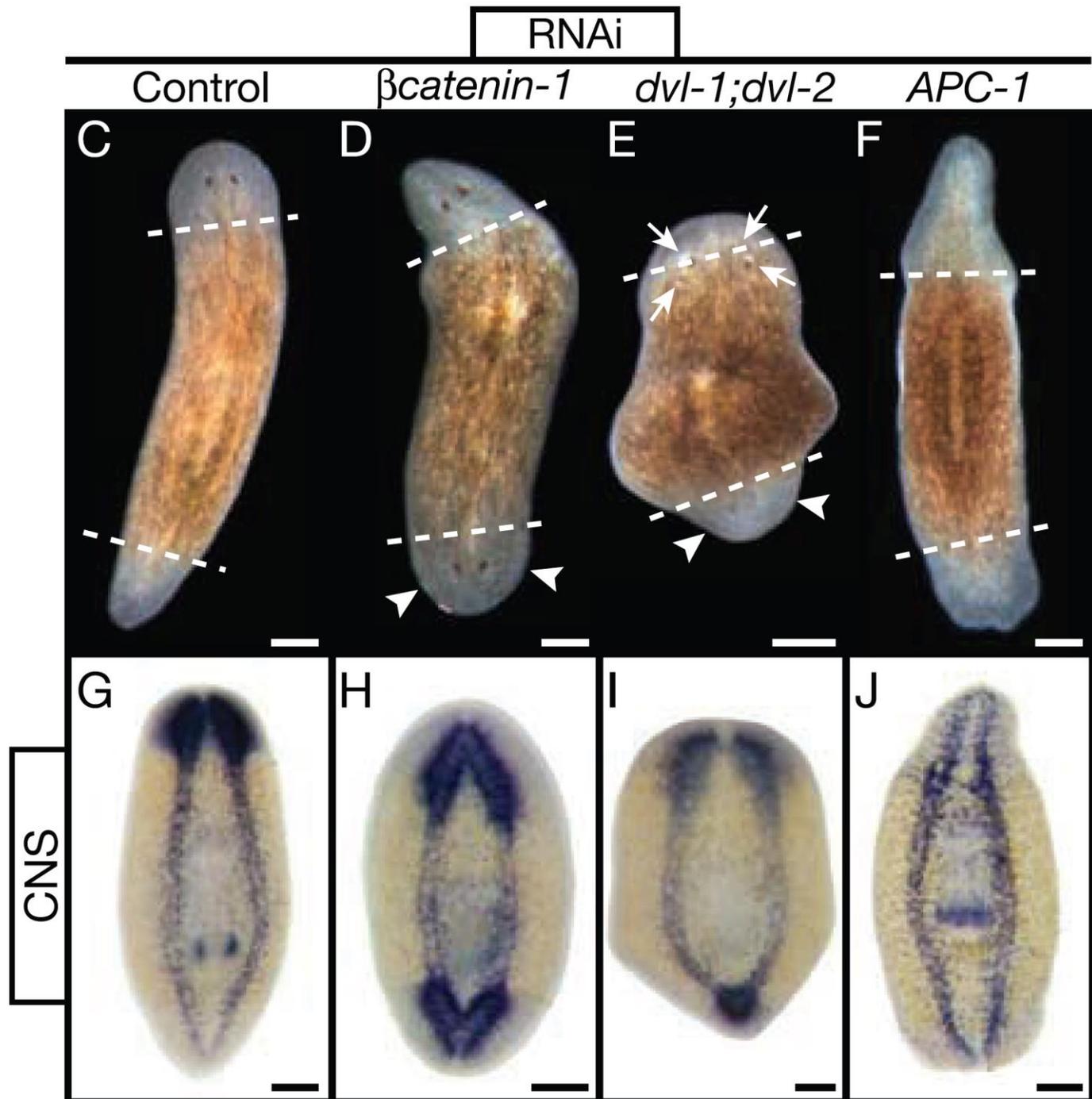


FIGURE 11.27 Paracrine factor antagonists from the organizer are able to block specific paracrine factors to distinguish head from tail. (A) The pharyngeal endoderm that underlies the head secretes Dickkopf, Frzb, and Cerberus. Dickkopf and Frzb block Wnt proteins; Cerberus blocks Wnts, Nodal-related proteins, and BMPs. The prechordal plate secretes the Wnt blockers Dickkopf and Frzb, as well as BMP blockers Chordin and Noggin. The notochord contains the BMP blockers Chordin, Noggin, and Follistatin but does not secrete Wnt blockers. Insulin-like growth factor (IGF) from the head endomesoderm probably acts at the junction of the notochord and prechordal mesoderm. (B) Summary of paracrine antagonist function in the ectoderm. Brain formation requires inhibiting both the Wnt and BMP pathways. Spinal cord neurons are produced when Wnt functions without the presence of BMPs. Epidermis is formed when both the Wnt and BMP pathways are operating.

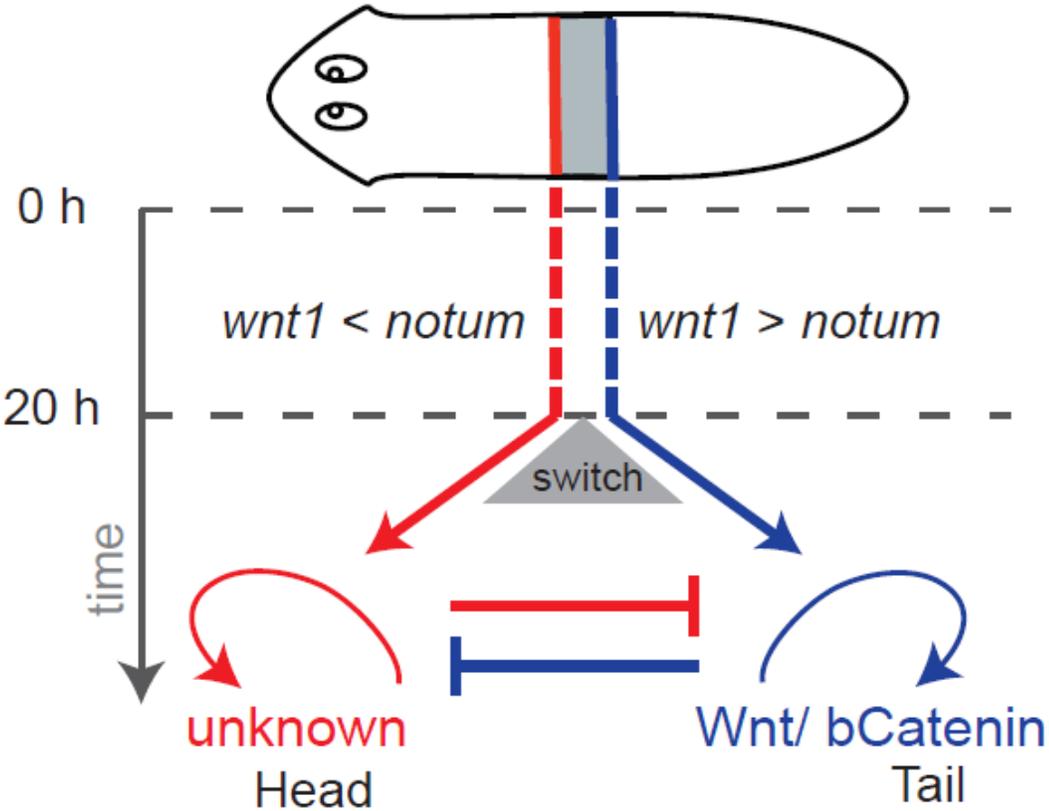
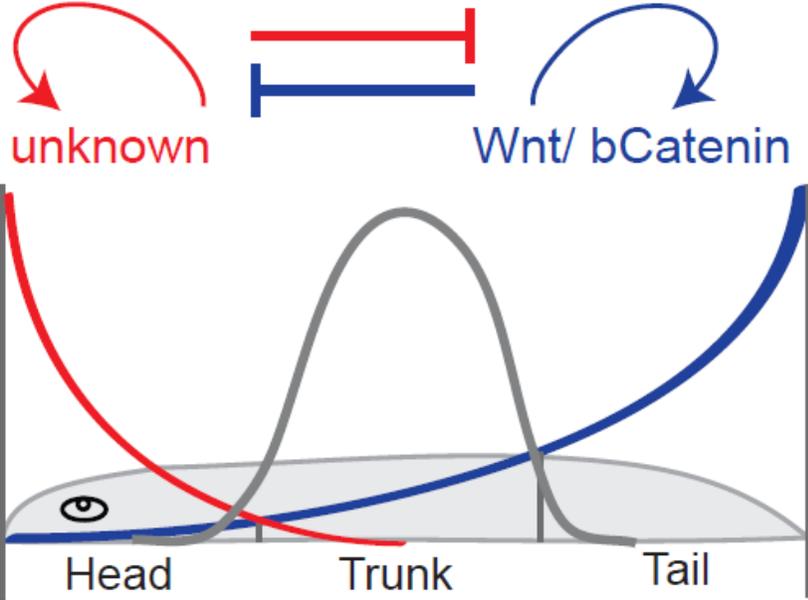


Species and system	Wild-type	Decrease in Wnt signaling activity	Increase in Wnt signaling activity
A Mouse head-tail patterning			
B Chick lateral mesoderm			
C Frog neurectoderm			
D Zebrafish head-tail patterning			
E Planarian regeneration			
F Nemertean animal-vegetal patterning			
G <i>C. elegans</i> axial asymmetric cell division			
H Cnidarian oral-aboral patterning			



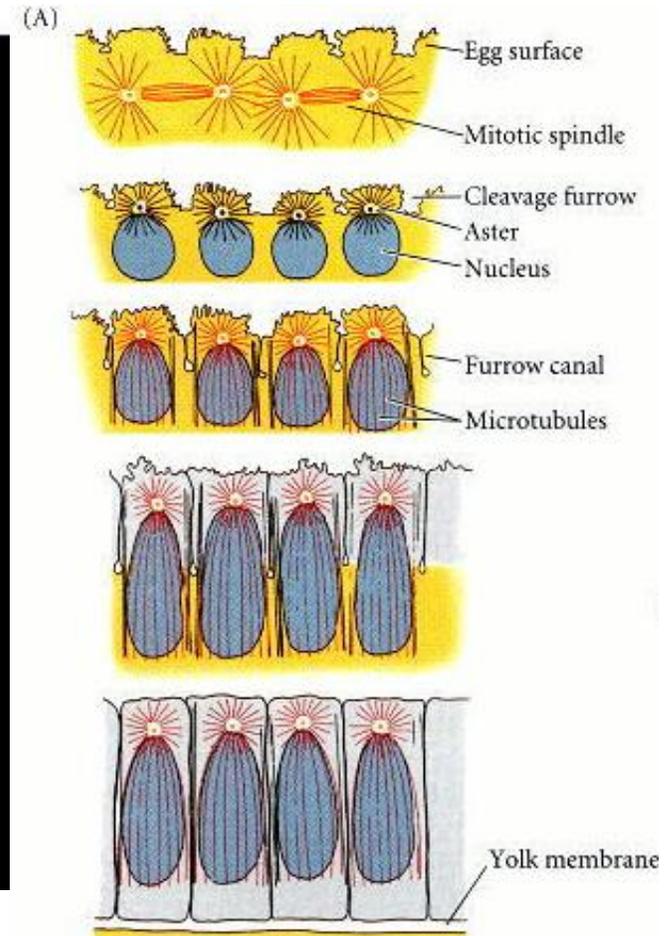
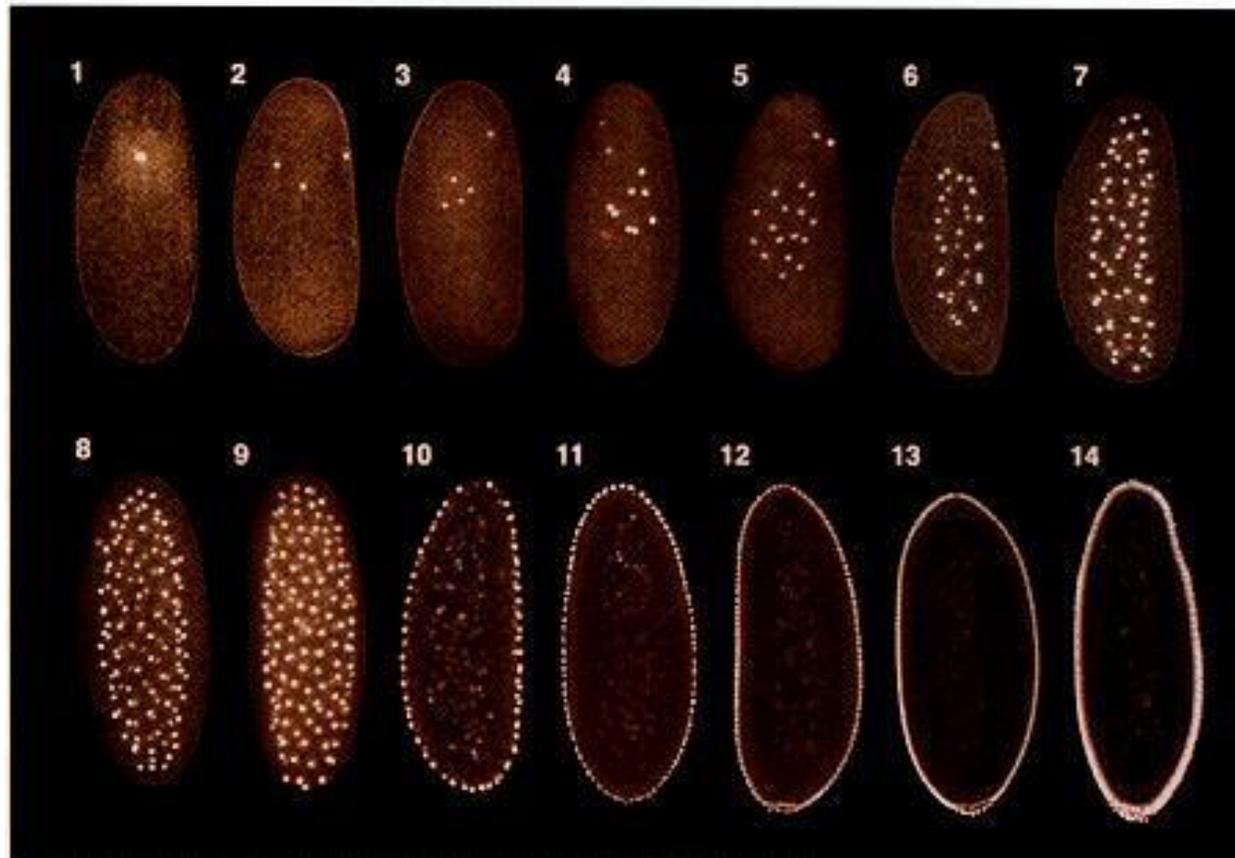
Regenerating planarian

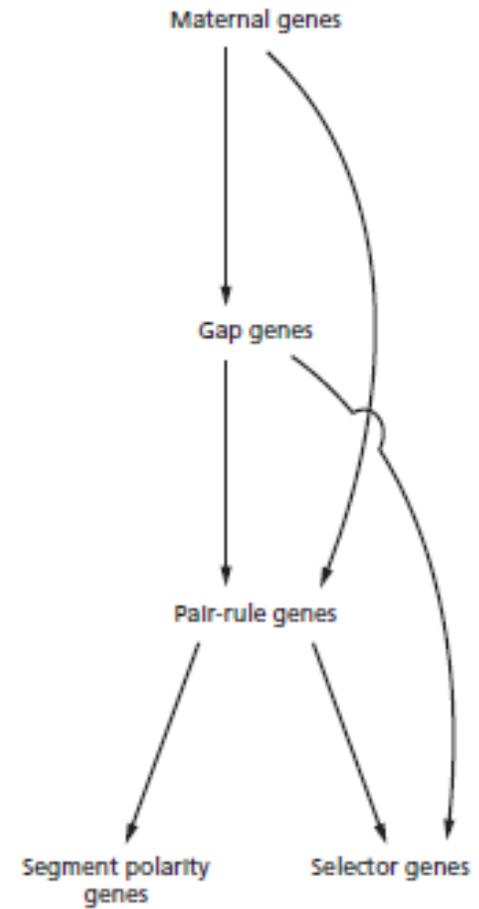
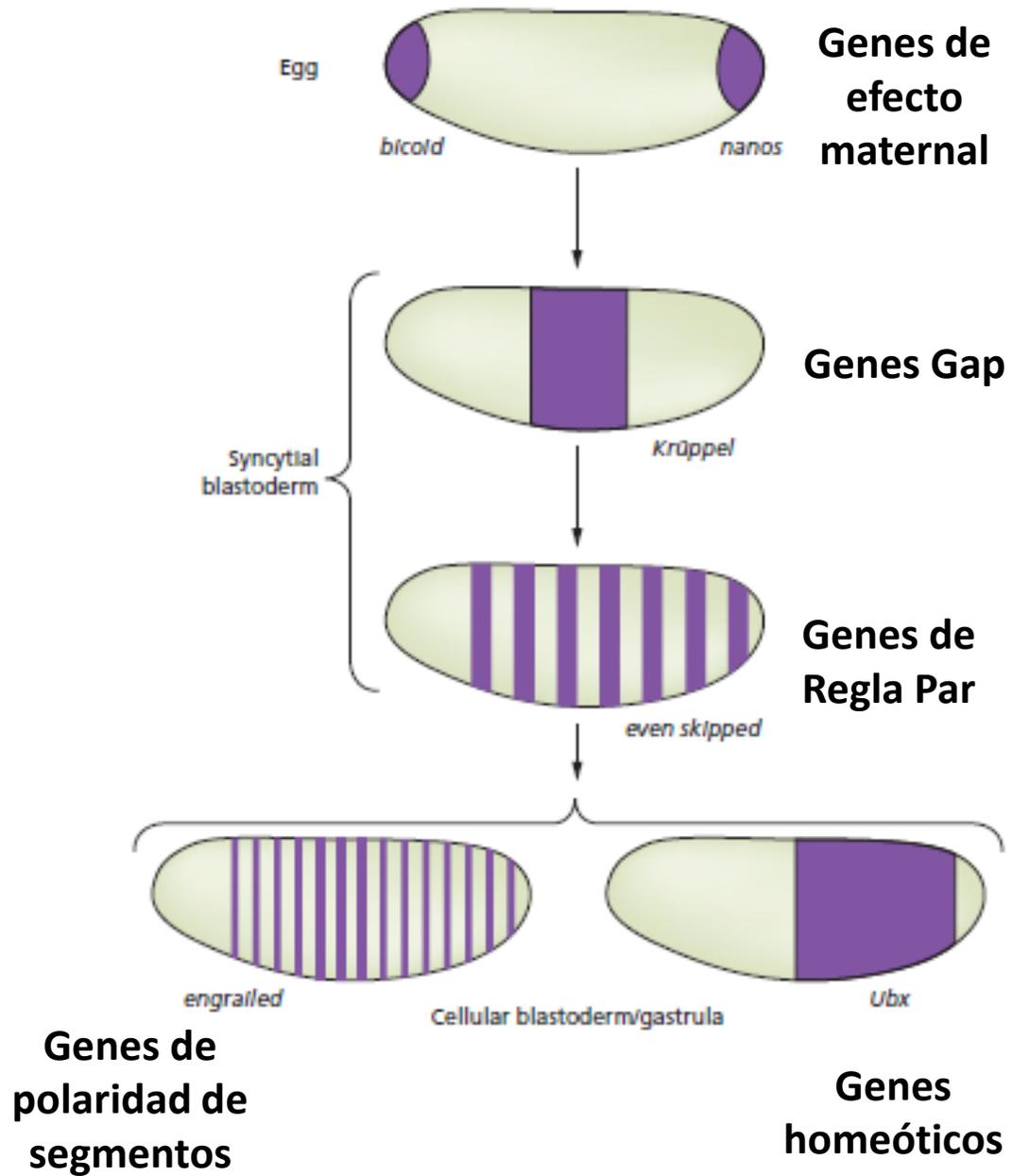
Intact planarian



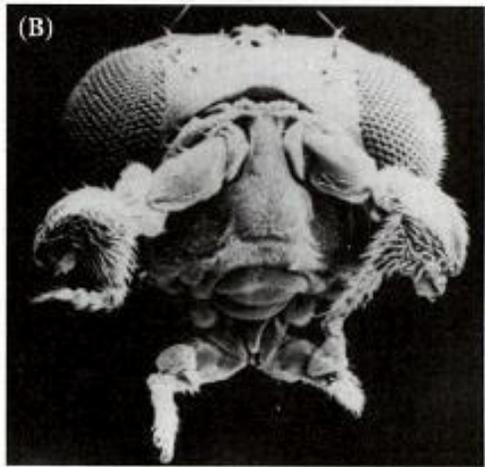
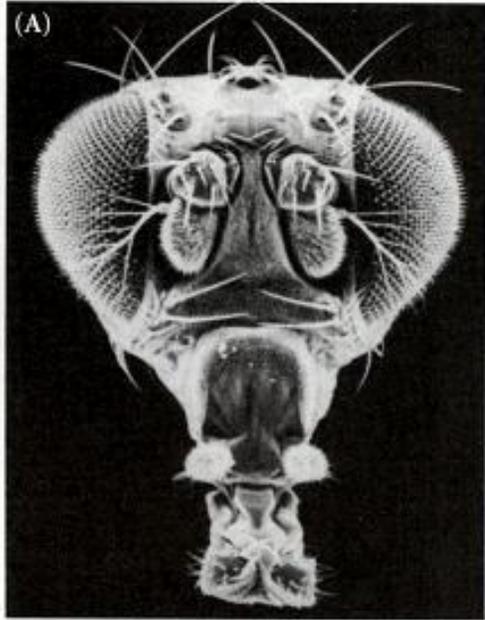
Cascadas regulatorias en el eje AP de *Drosophila*

Organización sincitial del embrión temprano de *Drosophila*





Mutaciones homeóóticas

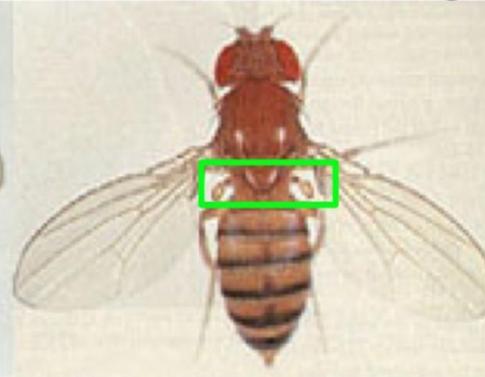


antenapedia

Ubx
loss of function



wild type

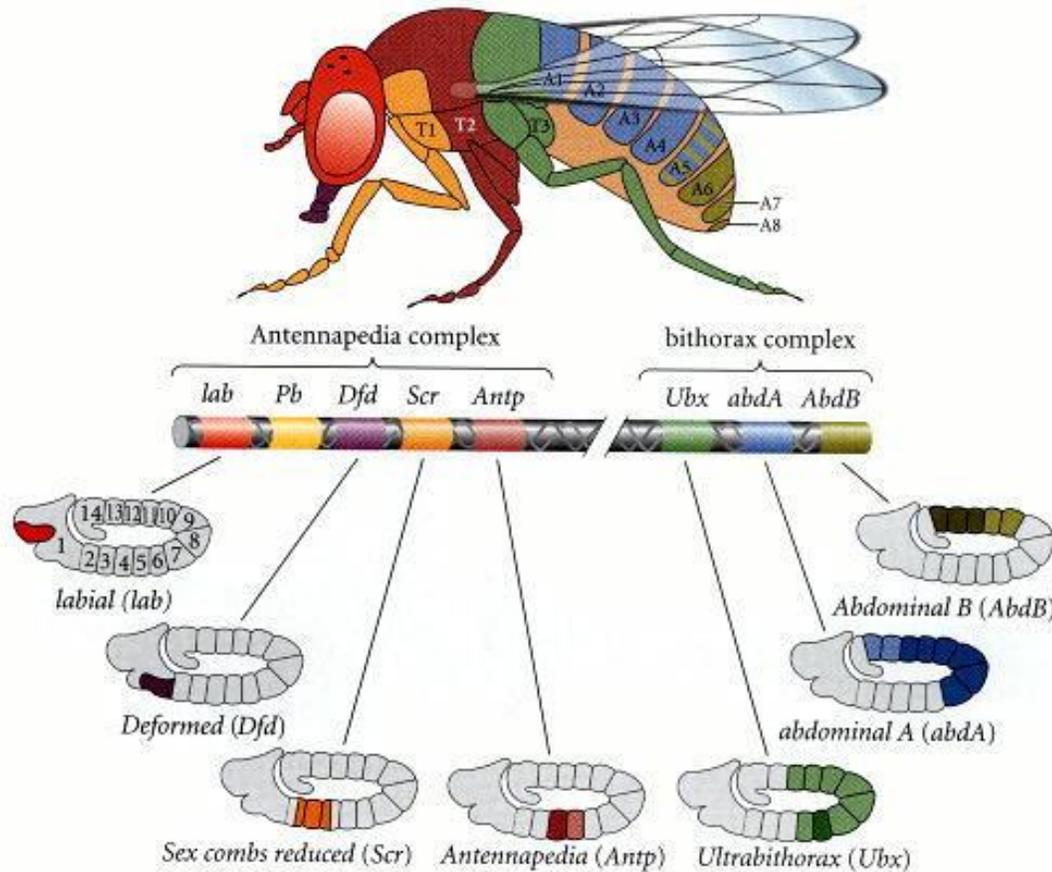


Ubx
gain of function



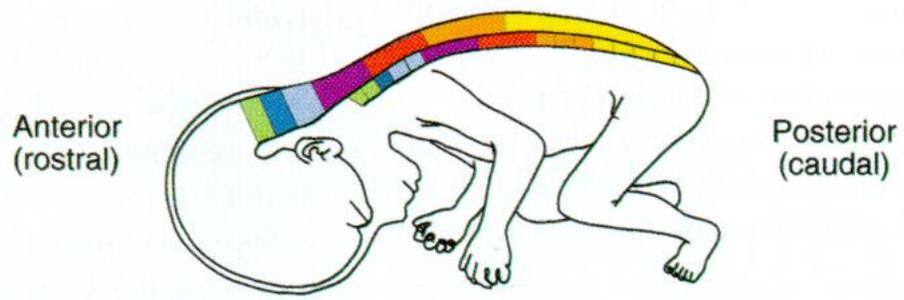
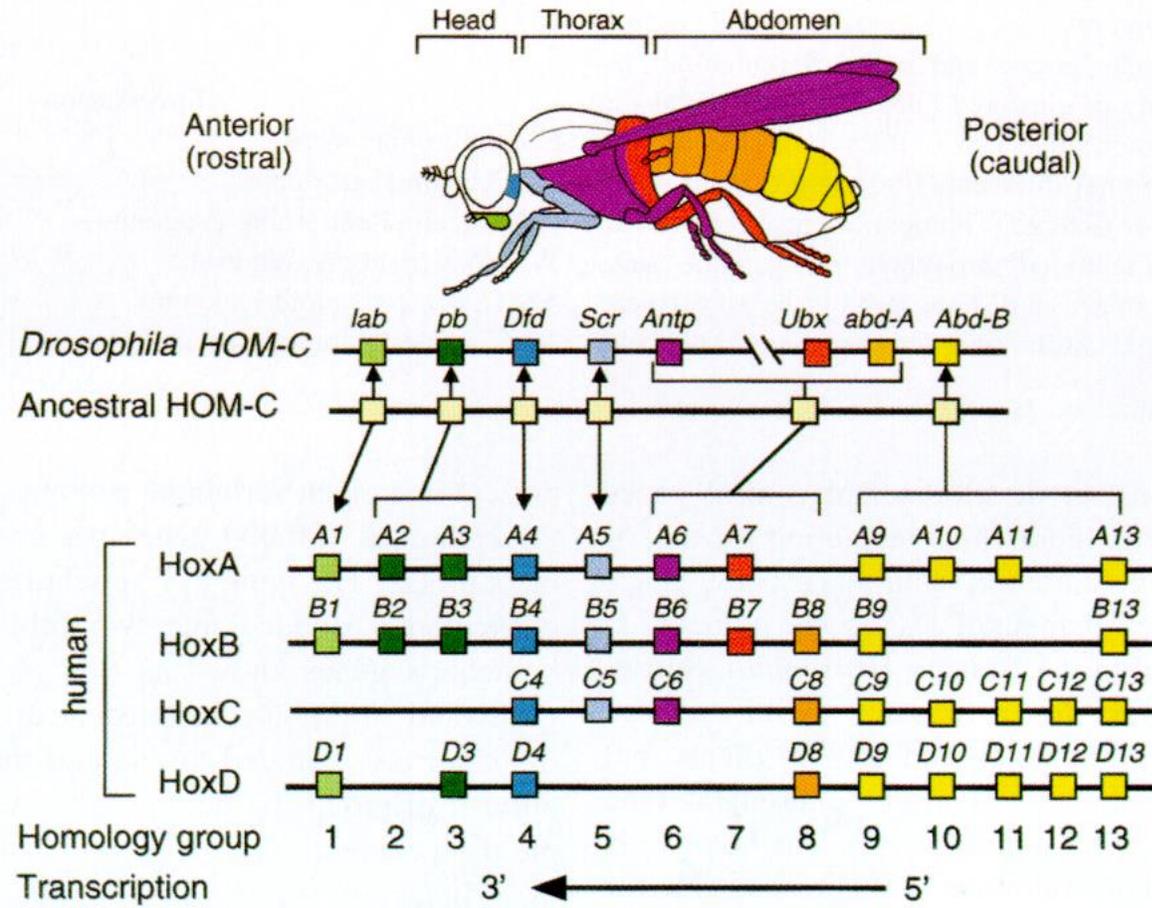
ultrabithorax

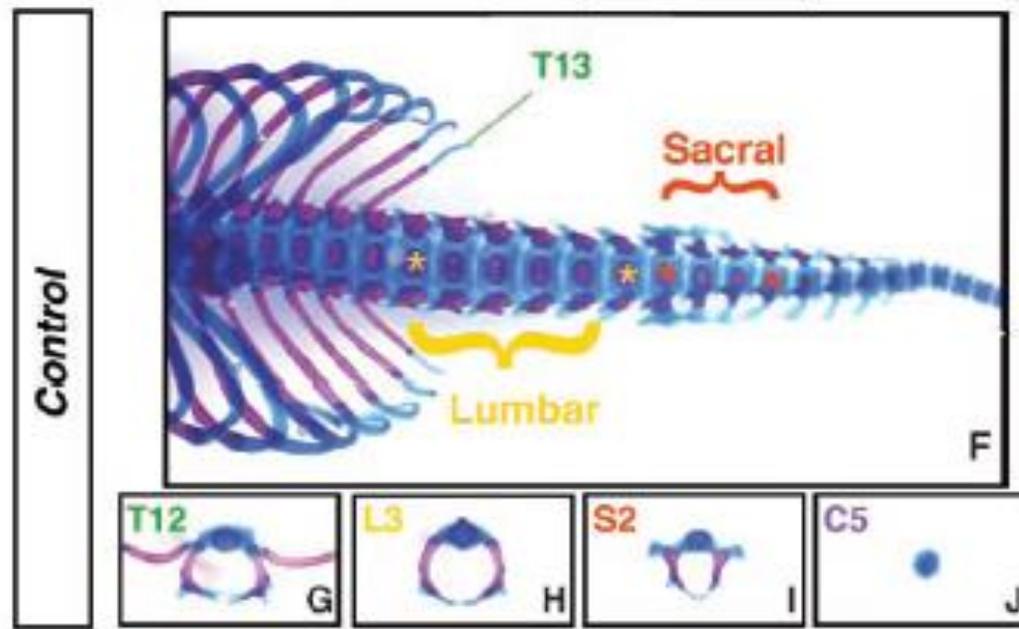
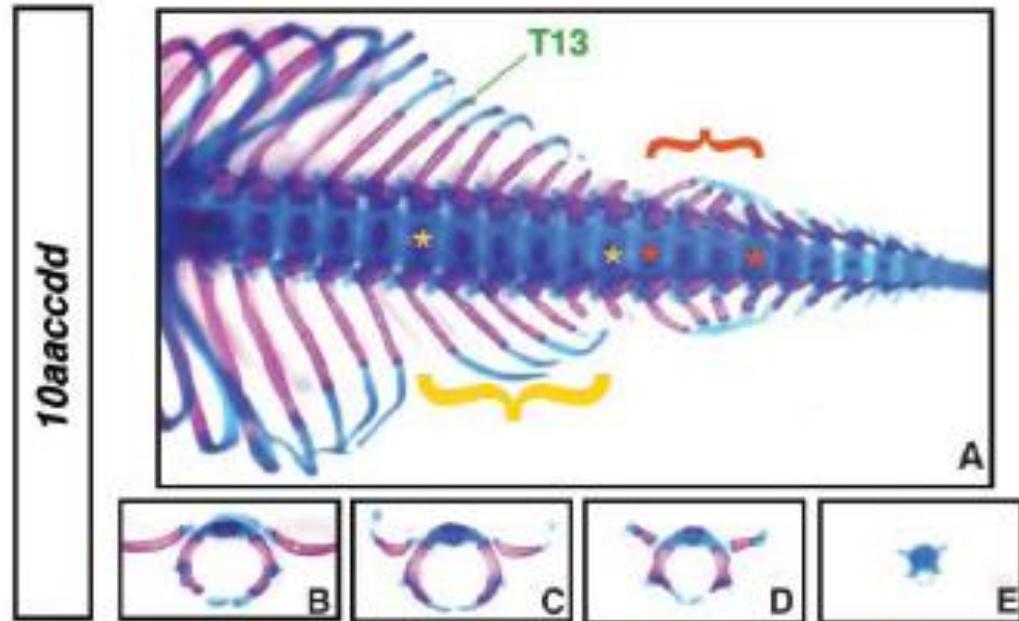
Mutaciones homeóticas



Complejo de genes Hox

- Factores de transcripción con homeodominio
- Especifican identidad A-P de segmentos
- Agrupados en el genoma (Cluster)
- Colinearidad entre posición en el complejo y expresión en el embrión!





Dominios de cromatina
Enhancers compartidos

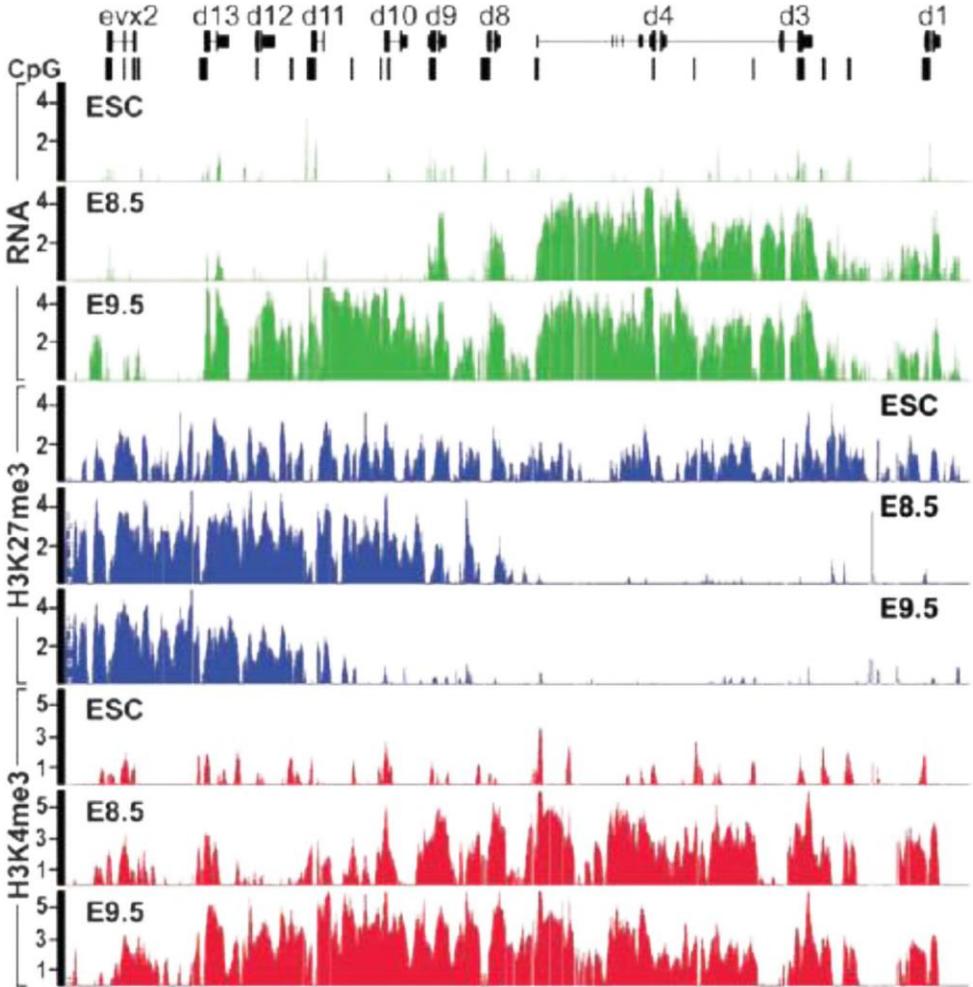
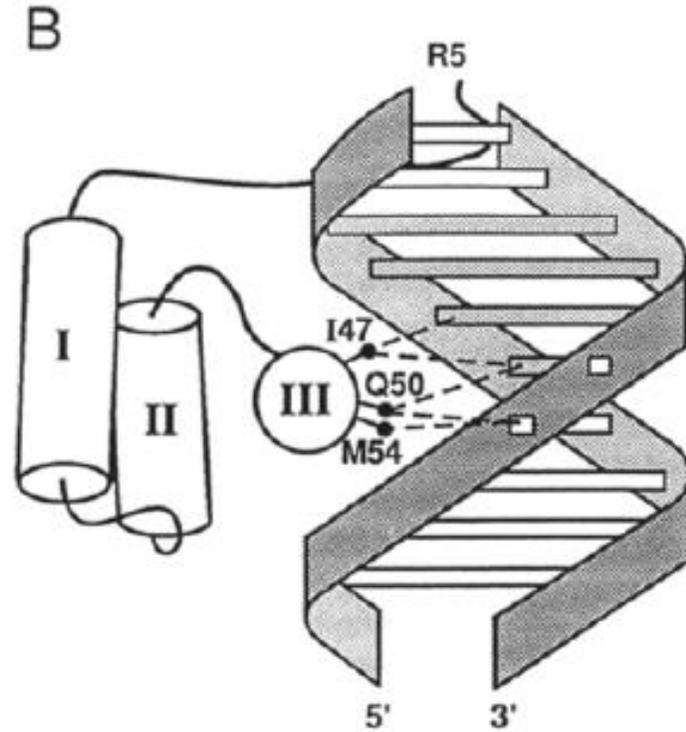
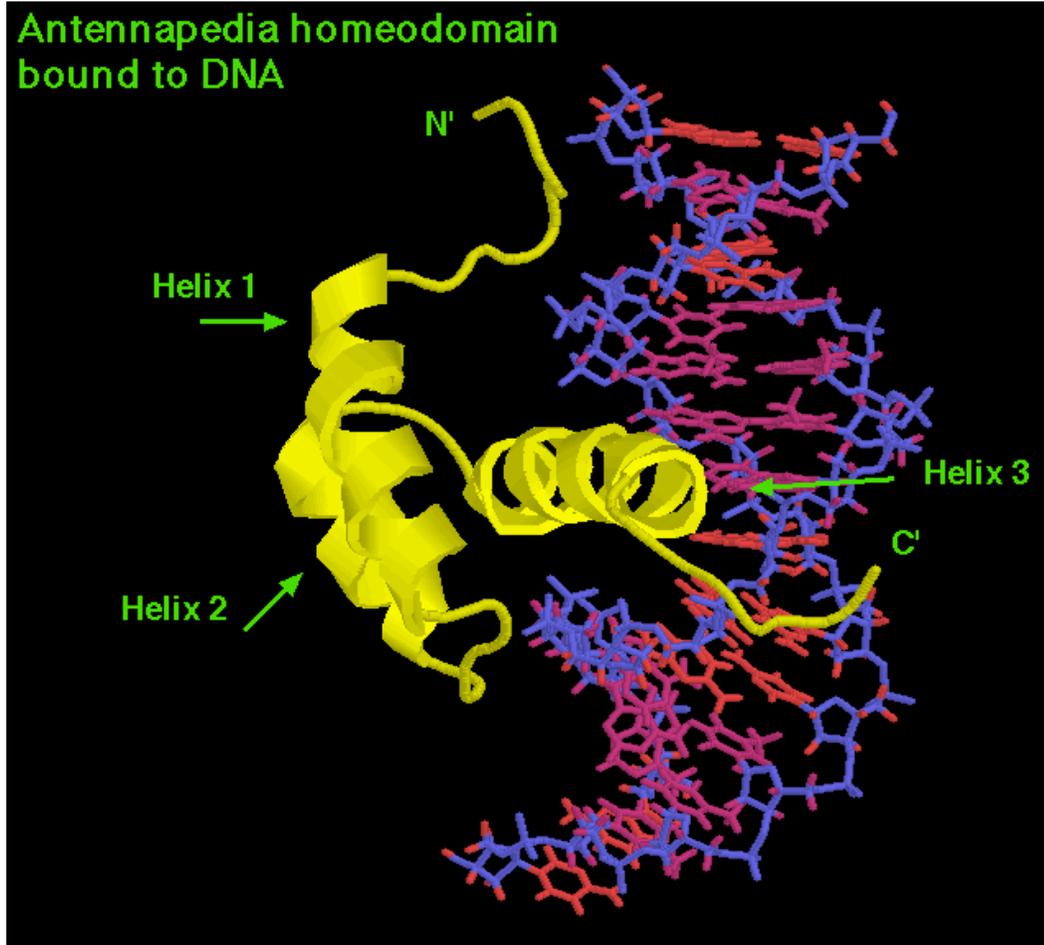
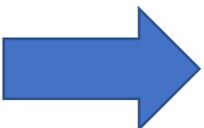
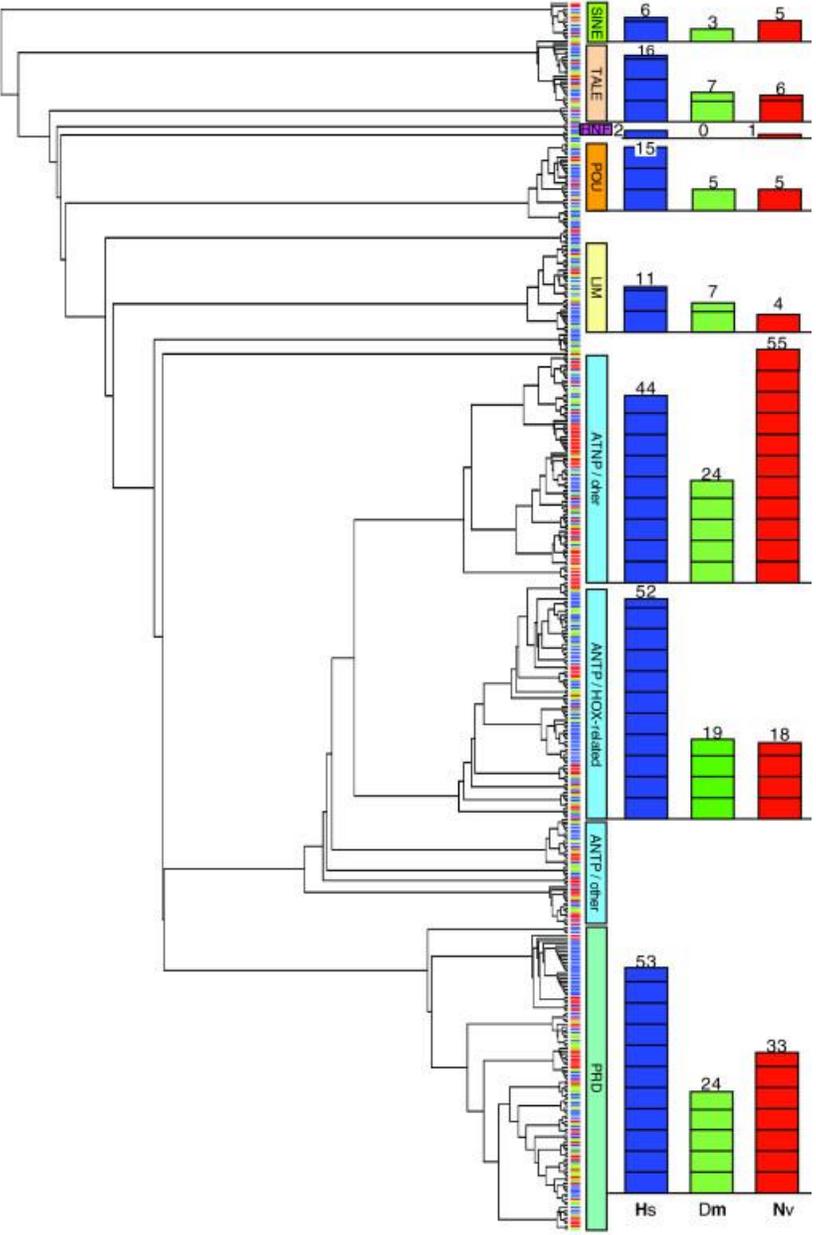


Fig 6. Chromatin marks and Hox expression in tailbuds from 8.5 day and 9.5 day mouse embryos. Over this period of development more posterior tissues arise within the tailbud and, correspondingly, more posterior Hox genes (Hoxd8 to d13) become expressed. There is accompanying loss of H3K27me3 and gain of H3K4me3 marks. H3K27me3 marks, in particular, suggest evidence of a single boundary between active and inactive Hox genes, and the position of this boundary in the cluster has shifted in the more posterior (older) tailbud. Embryo stem cells (ESC) are used to represent an earlier Hox-inactive stage. They show ubiquitous presence of both H3K27me3 and H3K4me3 marks, characteristic of bivalent chromatin, but at levels less than those reached after Hox gene expression. ESC, embryo stem cells. From Soshnikova and Duboule, 2009. Reprinted with permission from AAAS.

El homeodominio



Expansión de proteínas con homeodominio en Animales

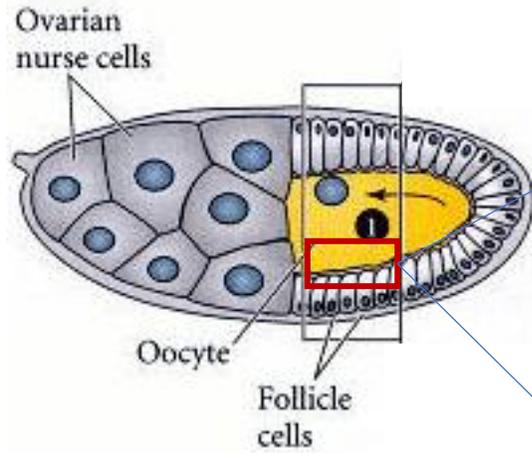


(Hox son sólo un grupo entre las proteínas con homeodominio)

Homo sapiens
Drosophila
Nematostella (Cnidaria)

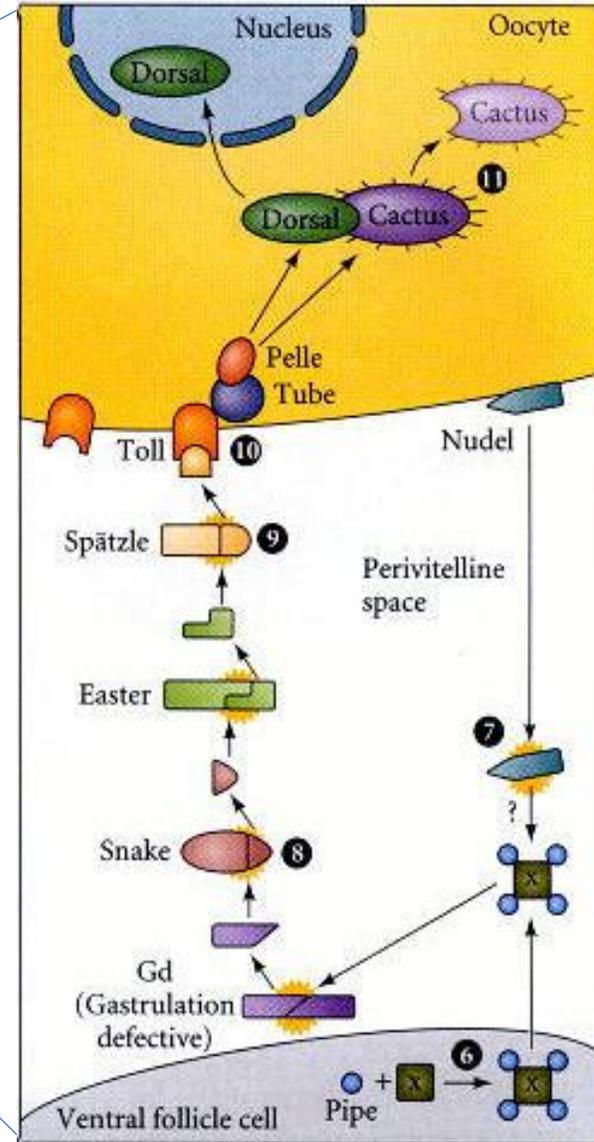
Eje Dorso-Ventral

Establecimiento del eje dorso-ventral en *Drosophila*

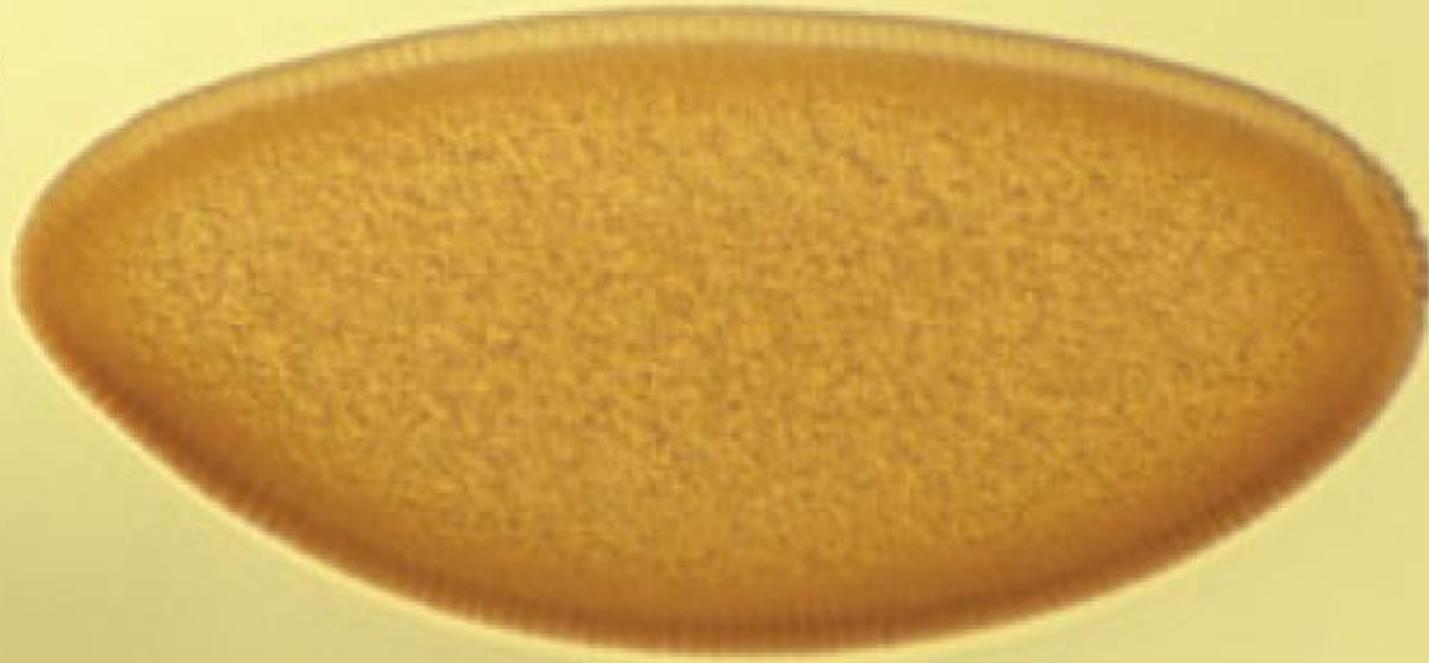


Cascada de proteólisis extracelular iniciada por células foliculares ventrales activa al ligando Spätzle

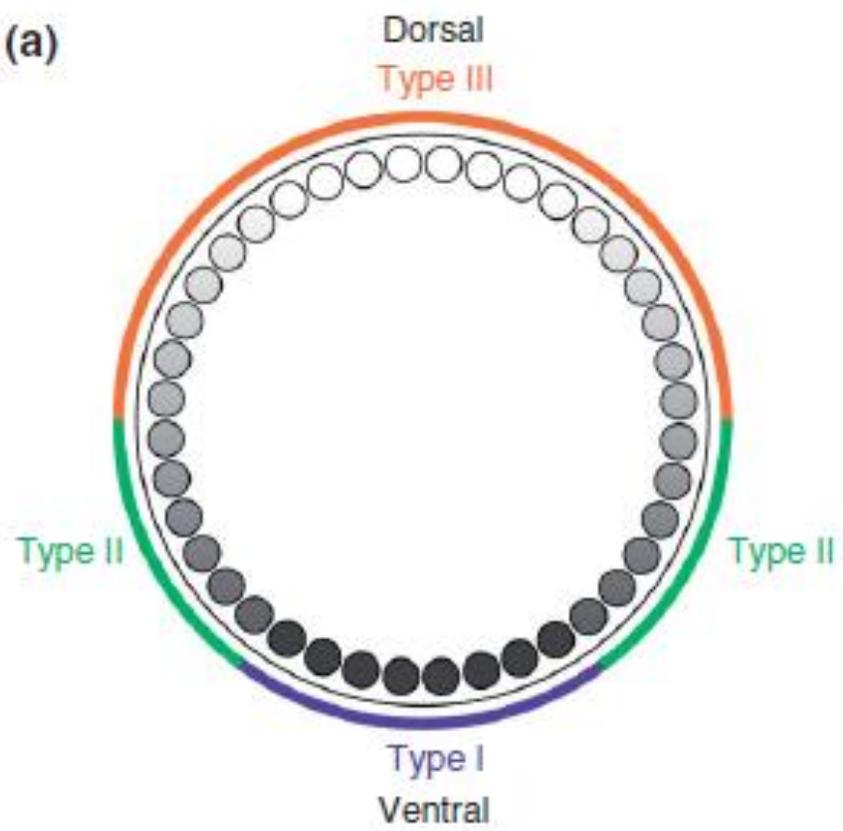
Activación del receptor Toll y del factor de transcripción Dorsal



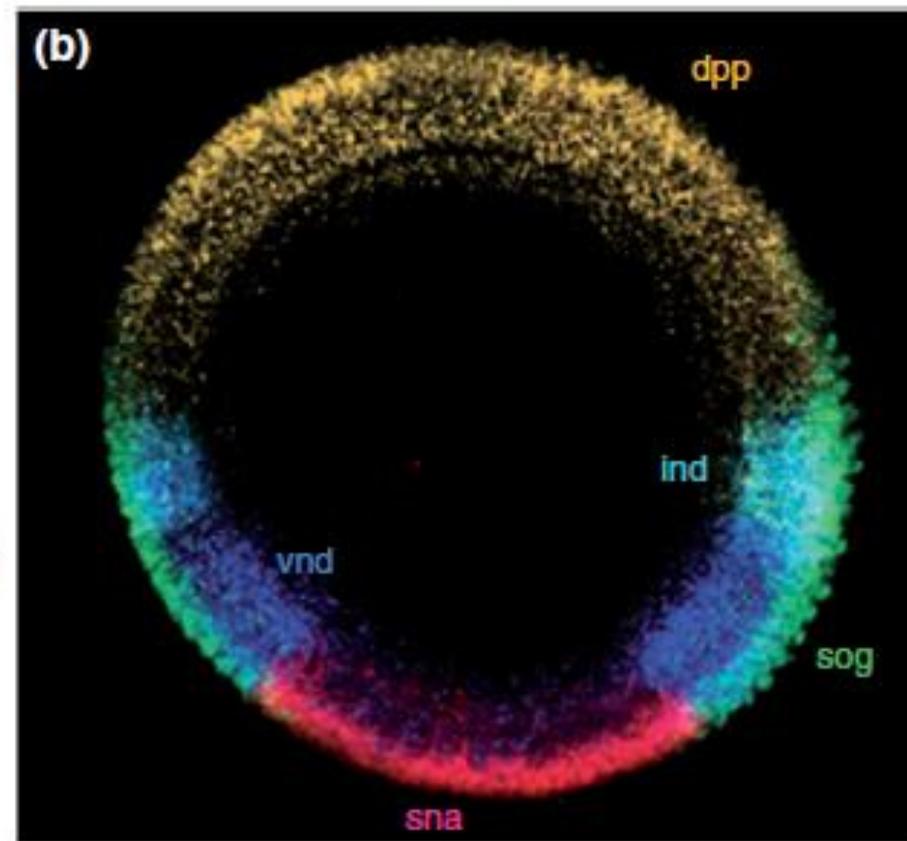
(e)

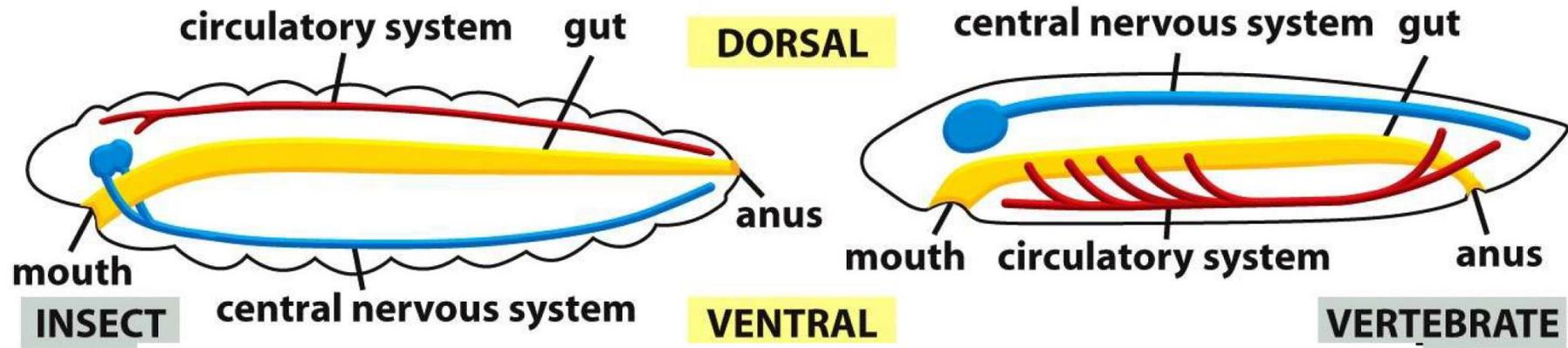


(a)



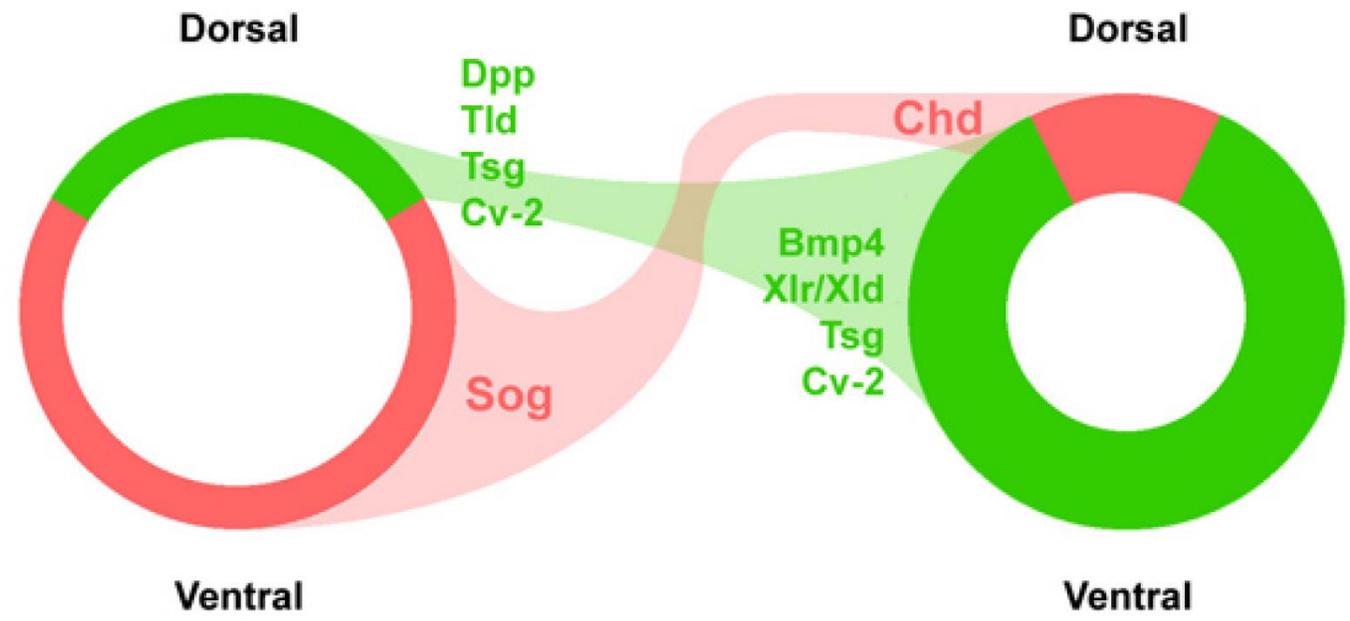
(b)





Drosophila

Xenopus



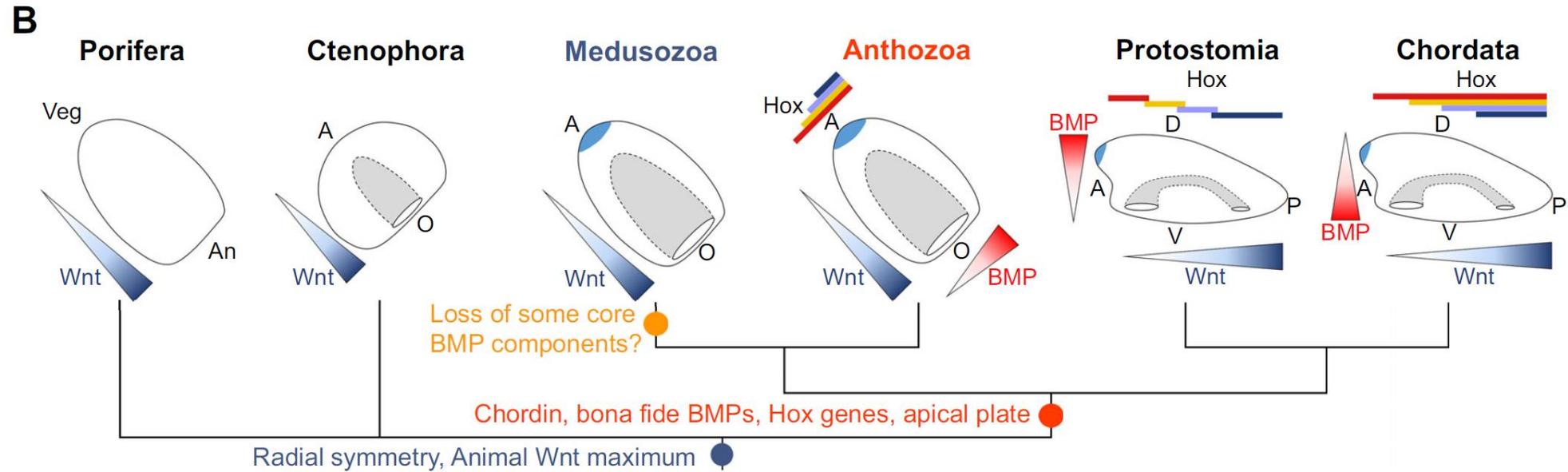
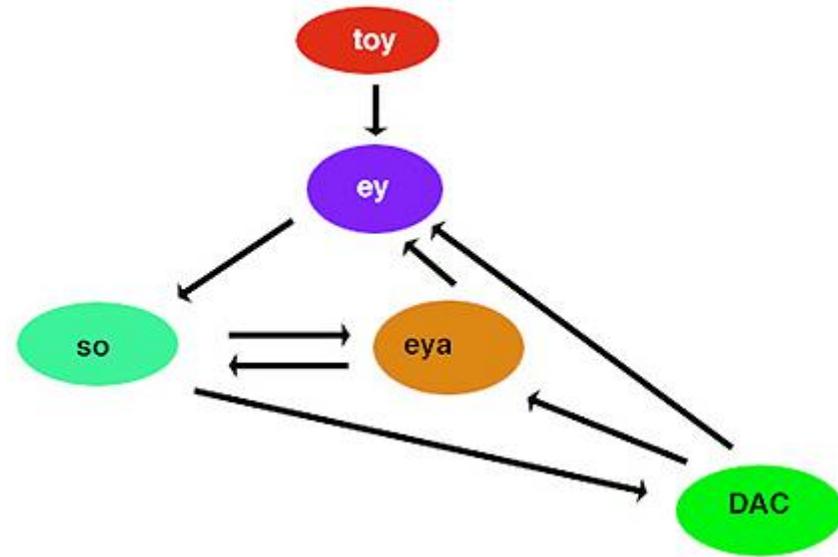


Fig. 5. The distribution of selected molecular traits amongst animals. (A) Table summarizing the presence or absence of several key regulatory components in the genomes and transcriptomes of Porifera, Ctenophora, Cnidaria and Bilateria. (B) The direction of the Wnt and BMP signalling gradients, and the expression of Hox genes, throughout Metazoa.

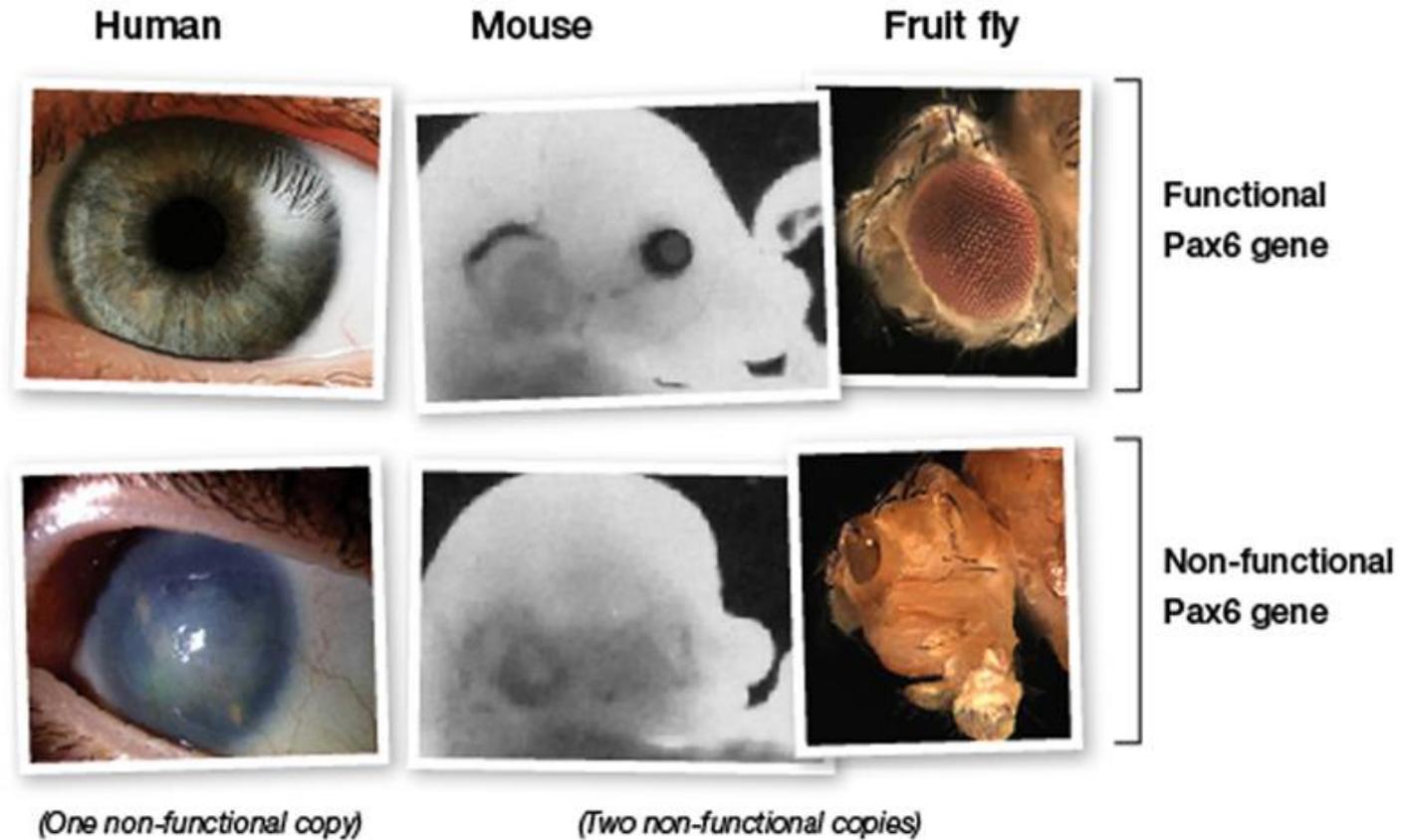
“Genes maestros”- Pax6 y el desarrollo del ojo



<http://www.ralf-dahm.com/>



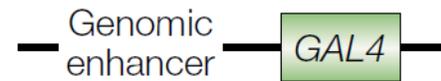
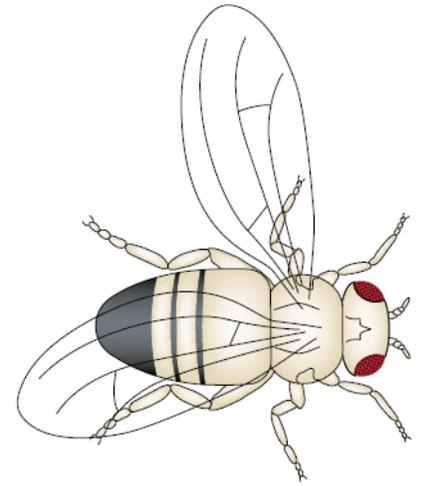
“Genes maestros”- Pax6 y el desarrollo del ojo



Box 2 | The **GAL4-UAS** system for directed gene expression

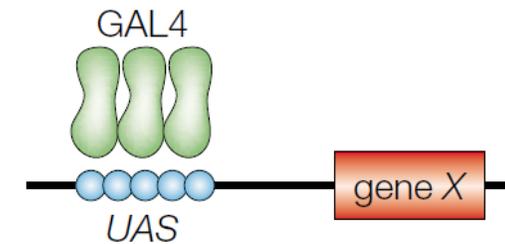
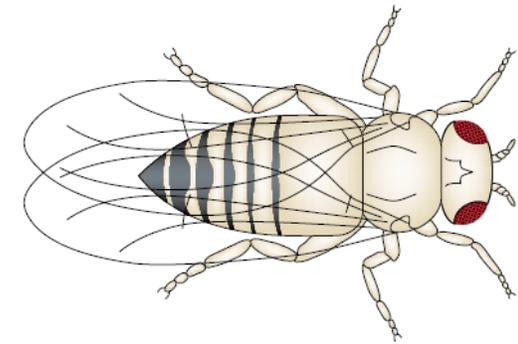
The yeast transcriptional activator Gal4 can be used to regulate gene expression in *Drosophila* by inserting the upstream activating sequence (*UAS*) to which it binds next to a gene of interest (*gene X*)⁹⁶. The *GAL4* gene has been inserted at random positions in the *Drosophila* genome to generate ‘enhancer-trap’ lines that express *GAL4* under the control of nearby genomic enhancers, and there is now a large collection of lines that express *GAL4* in a huge variety of cell-type and tissue-specific patterns⁹⁷. Therefore, the expression of *gene X* can be driven in any of these patterns by crossing the appropriate *GAL4* enhancer-trap line to flies that carry the *UAS-gene X* transgene. This system has been adapted to carry out genetic screens for genes that give phenotypes when misexpressed in a particular tissue (modular misexpression screens)⁷⁹.

Enhancer-trap *GAL4*



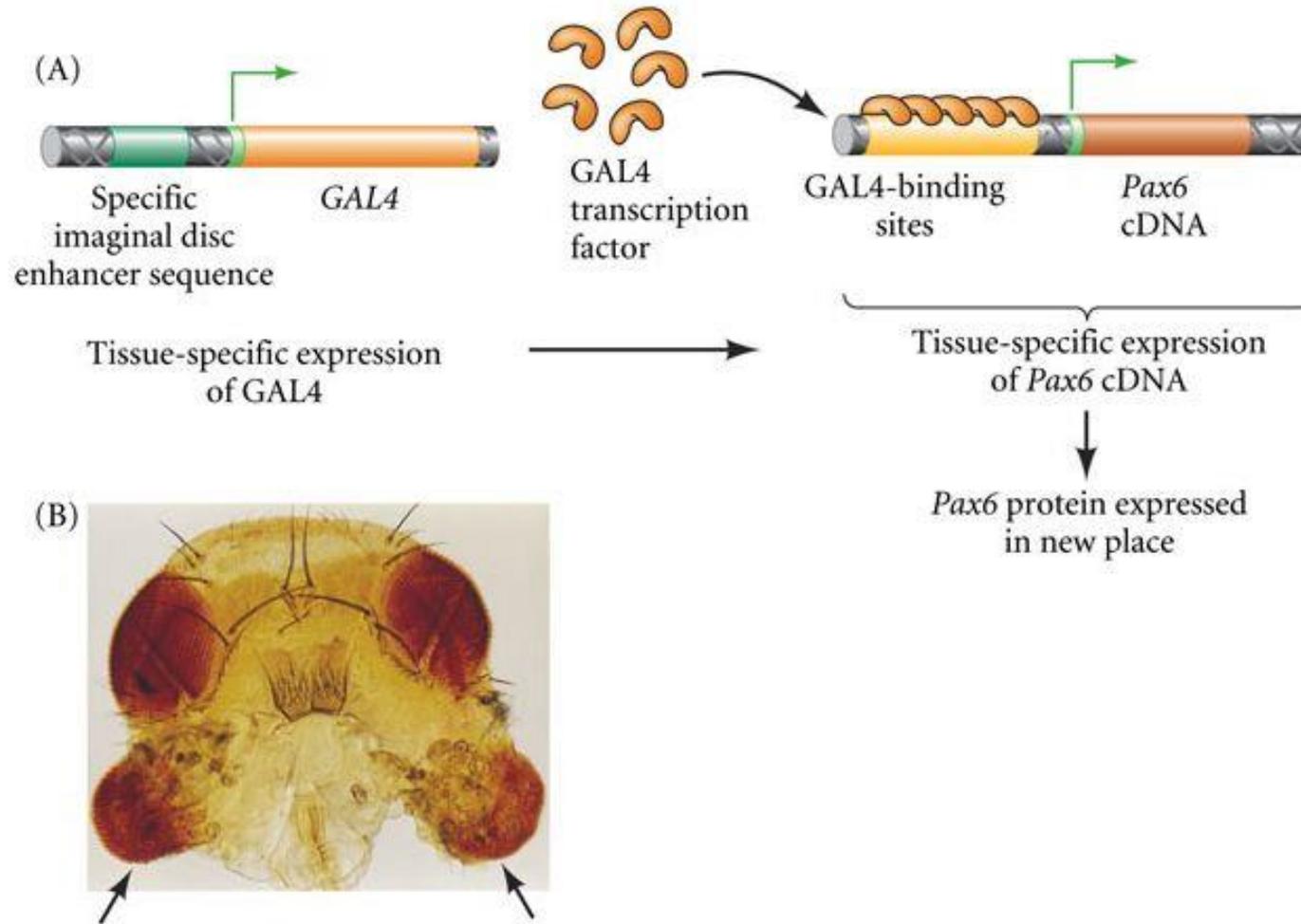
Tissue-specific expression of *GAL4*

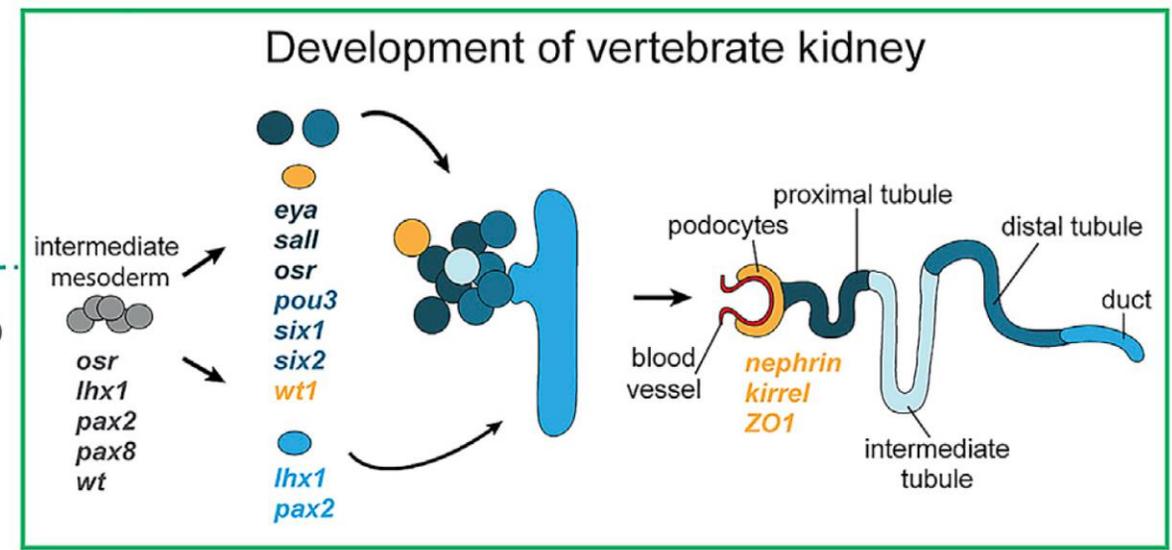
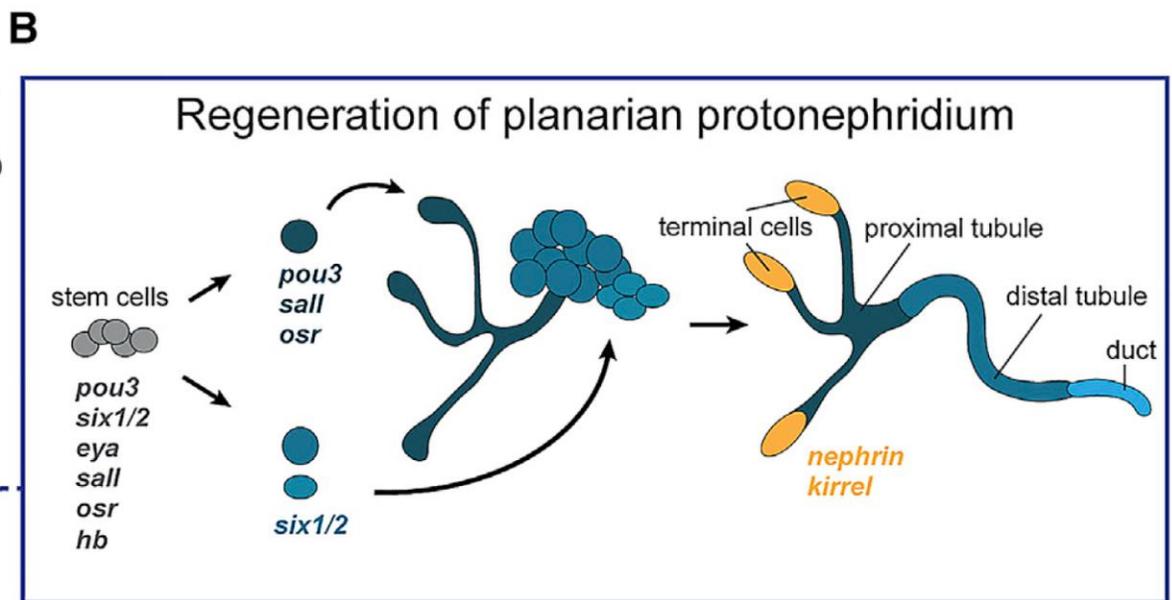
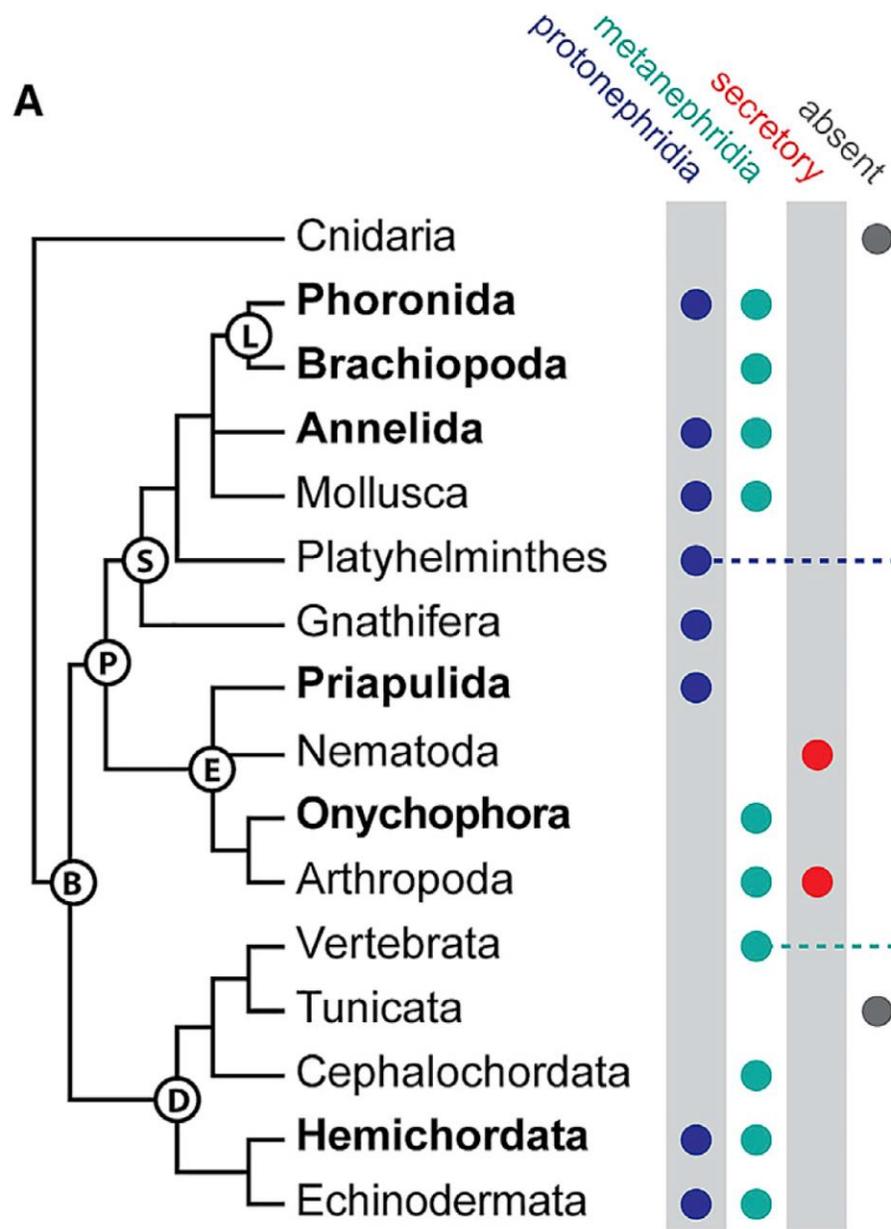
UAS-gene X



Transcriptional activation of *gene X*

“Genes maestros”- Pax6 y el desarrollo del ojo





El “zootipo” y el antepasado común de los animales bilaterales (“urbilateria”)

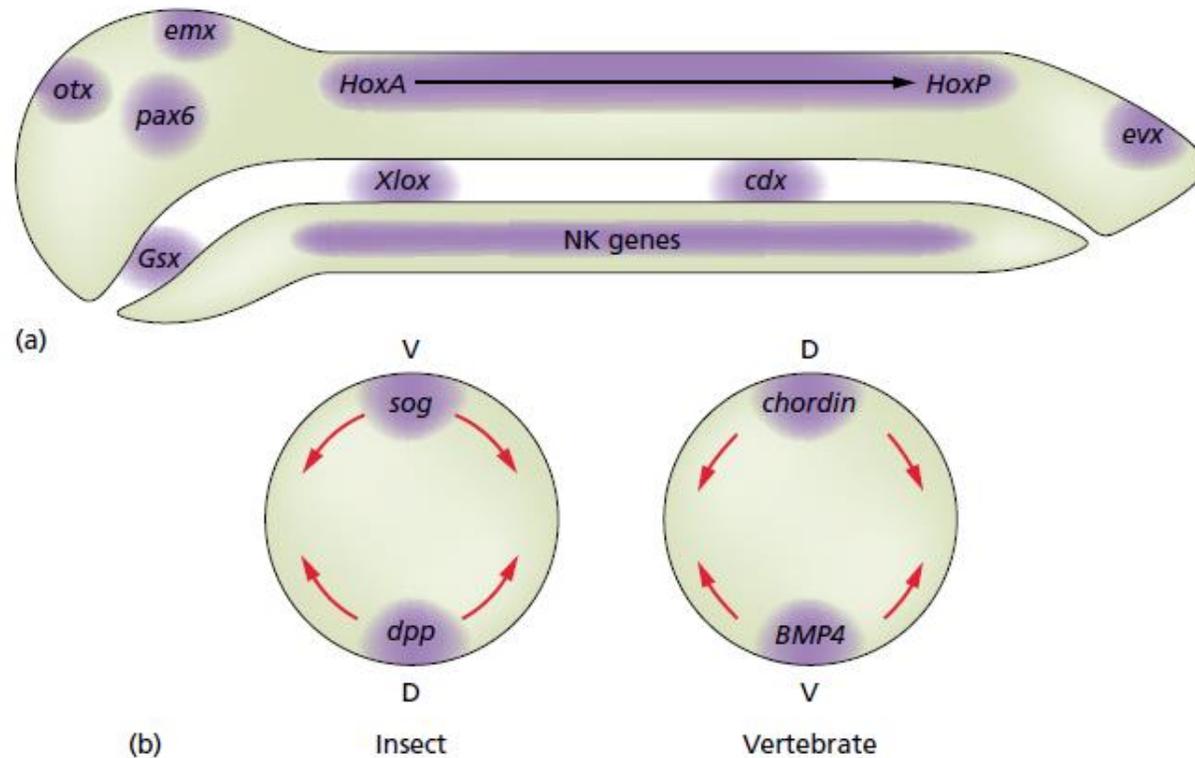
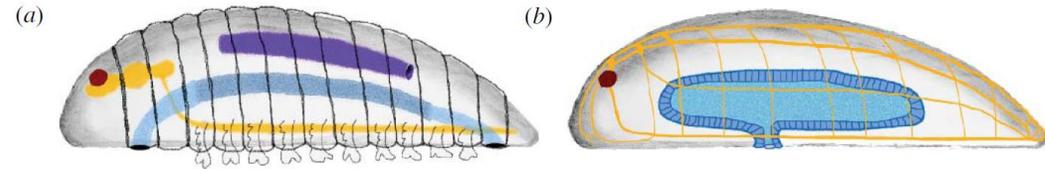


Fig. 20.6 The “zootype,” or common constellation of expression domains for developmental genes found in the animal body plan. (a) Transcription factors active at the phylotypic stage. (b) Homologous dorsoventral signaling systems in insects and vertebrates.



(c) character	genes	old	new
AP patterning	<i>HOX</i>		
DV patterning	<i>TGFβ/BMP 2/4</i> <i>sog/dpp</i>		
posterior patterning	<i>evx, cdx</i>		
central nervous system	<i>otx, emx, six3/6, HOX1</i>		
photoreception 'eyes'	<i>PAX6, RX, opsin</i>		
heart	<i>tinman</i>		
segmentation, segmentation clock	<i>hairy, engrailed, notch/delta</i>		
regionalized through gut	<i>HNFβ 3, GATA factor</i> <i>goosecoid, brachyury</i>		
appendages	<i>Distal-less/DLX</i>		

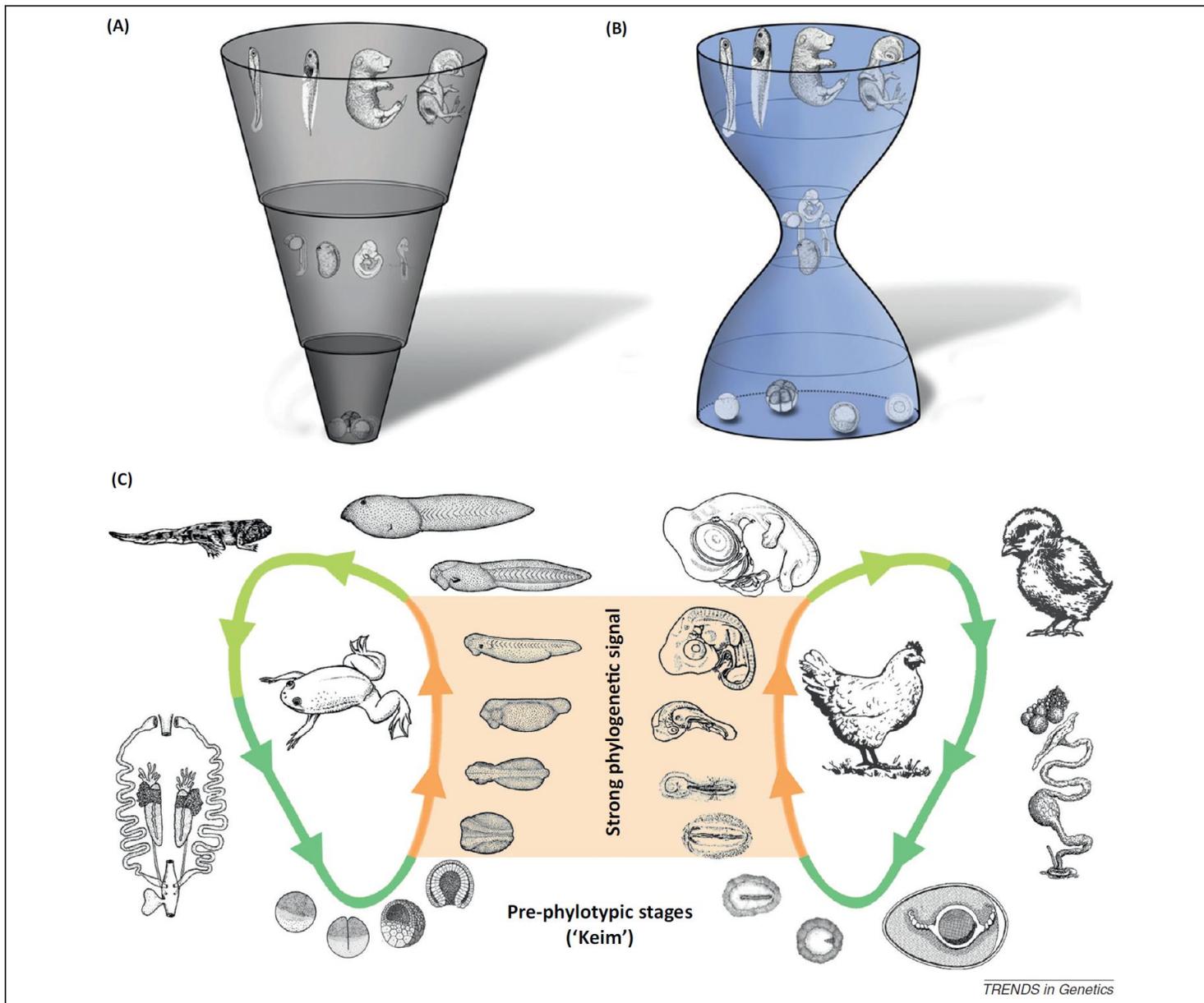


Figure 6. Different views of developmental evolution: the ‘funnel’ model (A), which suggests early conservation and the ‘hourglass’ model (B), which limits conservation only to the phylotypic stage [76,88]. Karl von Baer’s original observations suggested a high degree of parallelism in biological complexity and developmental changes for the mid- to late-stage embryos (C). Orange indicates embryonic development from the pharyngula stage to hatching, light green indicates postnatal development, and dark green indicates the adult stage.