# **Genetic Alterations Detected in Cell-Free DNA Are Associated With Enzalutamide and Abiraterone Resistance in Castration-Resistant Prostate Cancer**

Samantha Torquato, PhD<sup>1</sup>; Aparna Pallavajjala, MS<sup>1</sup>; Alexa Goldstein, MS<sup>1</sup>; Patricia Valda Toro, MS<sup>1</sup>; John L. Silberstein, MS<sup>1</sup>; Justin Lee<sup>1</sup>; Mary Nakazawa, MS<sup>1</sup>; Ian Waters<sup>1</sup>; David Chu, PhD<sup>1</sup>; Daniel Shinn<sup>1</sup>; Taylor Groginski<sup>1</sup>; Robert M. Hughes, PhD<sup>1</sup>; Brian W. Simons, DVM, PhD<sup>1</sup>; Hamda Khan, MS<sup>1</sup>; Zhaoyong Feng, MS<sup>1</sup>; Michael A. Carducci, MD, FACP<sup>1</sup>; Channing J. Paller, MD<sup>1</sup>; Samuel R. Denmeade, MD<sup>1</sup>; Bruce Kressel, MD<sup>1</sup>; Mario A. Eisenberger, MD<sup>1</sup>; Emmanuel S. Antonarakis, MBBCh<sup>1</sup>; Bruce J. Trock, PhD<sup>1</sup>; Ben H. Park, MD, PhD<sup>1,2</sup>; and Paula J. Hurley, PhD<sup>1</sup>

**PURPOSE** Androgen receptor (AR) gene alterations, including ligand-binding domain mutations and copy number (CN) gain, have yet to be fully established as predictive markers of resistance to enzalutamide and abiraterone in men with metastatic castration-resistant prostate cancer (mCRPC). The goal of this study was to validate AR gene alterations detected in cell-free DNA (cfDNA) as markers of enzalutamide and abiraterone resistance in patients with mCRPC.

**METHODS** Patients with mCRPC (N = 62) were prospectively enrolled between 2014 and 2018. Blood was collected before therapies—enzalutamide (n = 25), abiraterone (n = 35), or enzalutamide and abiraterone (n = 2) —and at disease progression. We used deep next-generation sequencing to analyze cfDNA for sequence variants and CN status in AR and 45 additional cancer-associated genes. Primary end points were prostatespecific antigen response, progression-free survival (PFS), and overall survival (OS).

**RESULTS** Elevated tumor-specific cfDNA (circulating tumor DNA) was associated with a worse prostate-specific antigen response (hazard ratio [HR], 3.17; 95% CI, 1.11 to 9.05; P = .031), PFS (HR, 1.76; 95% CI, 1.03 to 3.01; P = .039), and OS (HR, 2.92; 95% CI, 1.40 to 6.11; P = .004). AR ligand-binding domain missense mutations (HR, 2.51; 95% CI, 1.15 to 5.72; P = .020) were associated with a shorter PFS in multivariable models. AR CN gain was associated with a shorter PFS; however, significance was lost in multivariable modeling. Genetic alterations in tumor protein p53 (HR, 2.70; 95% Cl, 1.27 to 5.72; P = .009) and phosphoinositide 3-kinase pathway defects (HR, 2.62; 95% CI, 1.12 to 6.10; P = .026) were associated with a worse OS in multivariable models.

**CONCLUSION** These findings support the conclusion that high circulating tumor DNA burden is associated with worse outcomes to enzalutamide and abiraterone in men with mCRPC. Tumor protein p53 loss and phosphoinositide 3-kinase pathway defects were associated with worse OS in men with mCRPC. AR status associations with outcomes were not robust, and additional validation is needed.

#### JCO Precis Oncol. © 2019 by American Society of Clinical Oncology

Licensed under the Creative Commons Attribution 4.0 License

## ASSOCIATED CONTENT

**Data Supplement** 

Author affiliations and support information (if applicable) appear at the end of this article Accepted on January 25, 2019 and published at ascopubs.org/journal/ po on April 3, 2019: DOI https://doi.org/10.



1200/P0.18.00227

INTRODUCTION

Next-generation therapies that target the androgenandrogen receptor (AR) axis, such as abiraterone and enzalutamide, have improved survival outcomes for men with metastatic castration-resistant prostate cancer (mCRPC),<sup>1-4</sup> but both primary and acquired resistance to these drugs continue to be a substantial clinical challenge. Resistance mechanisms are not fully understood; however, some forms of resistance likely involve alterations to AR, including amplification and ligand-binding domain (LBD) missense mutations. Although rare in primary prostate cancers,<sup>5-7</sup> AR

gene alterations are highly prevalent in mCRPC.<sup>8-13</sup> Metastatic tissue biopsies as a sole means to detect and observe changes in AR status is impractical, and thus cell-free DNA (cfDNA) is gaining traction as a minimally invasive and easily obtainable tumor biopsy surrogate. Previous studies using cfDNA from the blood to evaluate the association of AR gene aberrations with resistance to abiraterone and enzalutamide are inclusive.<sup>14-17</sup> AR copy number (CN) gain<sup>18,19</sup> and/ or amplification<sup>20</sup> or detection of two or more AR mutations<sup>20</sup> have been associated with worse outcomes to such therapies as abiraterone and enzalutamide. In

## **ICO**<sup>®</sup> Precision Oncology

contrast, a recent study demonstrated that neither *AR* CN gain, nor *AR* LBD mutations, were significantly associated with time to progression on abiraterone and enzalutamide therapies in multivariable models.<sup>17</sup> Thus, the role of *AR* gene aberrations in mediating resistance to androgen–AR axis therapies has not been fully determined, and additional prospective studies are needed for clinical validation.

*AR* gene alterations are only detected in a subset of patients who have either primary or acquired resistance to androgen–AR therapies, thereby highlighting the need to determine other mechanisms that mediate resistance. The *AR* splice variant *AR*-V7 is associated with resistance to enzalutamide and abiraterone<sup>21-23</sup> and is also associated with increased *AR* CN.<sup>24</sup> In addition to *AR*, alterations in other genes, including tumor protein p53 (*TP53*), phosphatase and tensin homolog (*PTEN*), and breast cancer gene 2 (*BRCA2*), are enriched in lethal prostate cancer.<sup>8-11</sup> Studies support the idea that lineage plasticity from an ARdependent to an AR-independent state through loss of *TP53* and retinoblastoma-associated protein 1 (*RB1*) mediates resistance to AR-targeted therapies.<sup>25-28</sup>

Consistent with this, *TP53* defects have been shown to be associated with worse outcomes with abiraterone and enzalutamide therapies.<sup>17</sup> The role of *BRCA2* and other homology-directed repair (HDR) genes in mediating resistance to enzalutamide and abiraterone has not been definitively determined. Although it has been reported that truncating mutations in *BRCA2* and ataxia-telangiectasia mutated (*ATM*) gene are associated with a shorter time to progression on enzalutamide and abiraterone,<sup>17</sup> other studies have indicated that HDR defects may be associated with a better response to therapy.<sup>29,30</sup>

The primary goal of this study was to determine whether AR CN gain and/or LBD mutations detected in cfDNA were associated with enzalutamide and abiraterone resistance in patients with mCRPC. The secondary goal was to determine if alterations in other genes that are enriched in lethal prostate cancer, including TP53, PTEN, and BRCA2, were associated with response to enzalutamide and abiraterone. In this study, high circulating tumor DNA (ctDNA) burden was significantly associated with prostate-specific antigen (PSA) response, progression-free survival (PFS), and overall survival (OS). AR LBD mutations were associated with a shorter PFS, whereas AR CN gain was associated with both a shorter PFS and worse OS, but lost significance in multivariable analyses. TP53 loss and defects in the phosphoinositide 3-kinase (PI3K) pathway were both associated with worse OS. Study limitations, including sample size and patient heterogeneity, necessitate larger and prospective validation of the association of plasma AR status with outcomes.

## METHODS

Patient information, study end points, sample collection, deep next-generation sequencing (NGS), sequence alignment and

analysis of variants, CN variation, estimation of ctDNA fraction, and statistical analyses are found in the Data Supplement.

## RESULTS

## Patient Cohort

Patient characteristics are listed in Table 1. PSA, PSA response, and PFS were not significantly different between patients on abiraterone and enzalutamide (Table 1 and Data Supplement). Approximately one quarter of patients had received prior abiraterone or enzalutamide. Prior abiraterone or enzalutamide exposure trended toward an association for worse outcomes, including PSA response (odds ratio [OR], 2.41; 95% CI, 0.74 to 7.93; P = .146), PFS (hazard ratio [HR], 1.17; 95% CI, 0.63 to 2.14; P = .620), and OS (HR, 1.51; 95% CI, 0.71 to 3.24; P = .284); however, these associations did not reach statistical significance (Tables 2 and 3 and Data Supplement). ClinVarannotated pathogenic or likely pathogenic missense mutations, truncating mutations, and/or CN alterations were detected in cfDNA from 89% of patients before therapy initiation and in 92% of patients at disease progression (Figs 1A-1D and Data Supplement).

## ctDNA

Total cfDNA concentration before therapy was associated with PSA (P = .002; Data Supplement). We used deep NGS to analyze cfDNA for CN variation and mutations in 46 cancer-associated genes (Data Supplement). Nearly all patients (61 of 62) had detectable CN variation(s) and/or mutation(s) with an allelic frequency above the 1% cutoff before therapy (Figs 1B and 1C). High ctDNA was detected in approximately 44% of patients before therapy (Fig 1C). Consistent with previous findings,<sup>14,17,20</sup> high ctDNA was significantly associated with a worse PSA response (OR, 3.17; 95% CI, 1.11 to 9.05; P = .031) by logistic regression analyses (Table 2). High ctDNA was associated with a significantly shorter median time to progression (14.0 weeks v 34.0 weeks; P = .022) and, using proportional hazards regression modeling, a shorter PFS (HR, 1.76; 95% CI, 1.03 to 3.01; P = .039; Table 3 and Fig 2A). High ctDNA was also significantly associated with a shorter median survival (62.7 weeks v 134.9 weeks; P = .003) and worse OS (HR, 2.92; 95% CI, 1.40 to 6.11; P = .004; Table 3 and Fig 3A). Other clinical variables, such as PSA, age, and visceral metastases, were not significantly associated with PSA response, PFS, or OS in univariable analyses (Tables 2 and 3 and Data Supplement).

## AR

Previous studies that evaluated associations between *AR* gene alterations, including CN gain and LBD missense mutations, with therapeutic outcomes are not definitive.<sup>14,15,17-20,31</sup> *AR* CN gain was detected in approximately one half of patients before therapy and at disease progression (Figs 1C and 1D and Data Supplement). *AR* CN gain was not

### TABLE 1. Patient Characteristics (N = 62)

Characteristic	Total Cohort (N = 62)	Abiraterone ( $n = 35$ )	Enzalutamide (n = $25$ )	Abiraterone Plus Enzalutamide (n = 2)
Age, years, median (range)	71.5 (41-90)	71 (51-90)	73 (41-90)	79 (70-87)
Race				
White	51 (82.3)	27 (77.1)	24 (96.0)	0 (0)
Black	7 (11.3)	6 (17.1)	0 (0)	1 (50)
Other	4 (6.5)	2 (5.7)	1 (4.0)	1 (50)
Local treatment of prostate cancer				
Radical prostatectomy	24 (38.7)	14 (40.0)	9 (36.0)	1 (50)
Radiation	9 (14.5)	3 (8.6)	5 (20.0)	1 (50)
Other	4 (6.5)	3 (8.6)	1 (4.0)	0 (0)
None	24 (38.7)	14 (40.0)	10 (40.0)	0 (0)
Not available	1 (1.6)	1 (2.9)	0 (0)	0 (0)
Gleason sum				
≤ 7	14 (22.6)	10 (28.6)	2 (8.0)	2 (100)
≥ 8	42 (67.7)	22 (62.9)	20 (80.0)	0 (0)
Not available	6 (9.7)	3 (8.6)	3 (12.0)	0 (0)
Prior treatment of metastatic prostate cancer				
Prior chemotherapy	13 (21.0)	5 (14.3)	8 (32.0)	0 (0)
Prior abiraterone	10 (16.1)	2 (5.7)	8 (32.0)	0 (0)
Prior enzalutamide	5 (8.1)	4 (11.4)	1 (4.0)	0 (0)
Median PSA, ng/mL (range)	19.3 (0.6-1966)	18.2 (0.6-1966)	18.9 (0.9-205.9)	102.6 (54.2-151.0)
Site of metastases				
Bone only	34 (54.8)	21 (60.0)	13 (52.0)	0 (0)
Visceral only	3 (4.8)	0 (0)	3 (12.0)	0 (0)
Lymph node only	4 (6.5)	1 (2.9)	3 (12.0)	0 (0)
Bone and visceral	3 (4.8)	1 (2.9)	1 (4.0)	1 (50)
Bone and lymph node	18 (29.0)	12 (34.3)	5 (20.0)	1 (50)
Study therapy				
Abiraterone plus prednisone	35 (56.5)	35 (100)	0 (0)	0 (0)
Enzalutamide	25 (40.3)	0 (0)	25 (100)	0 (0)
Abiraterone plus prednisone and enzalutamide	2 (3.2)	0 (0)	0 (0)	2 (100)
Median progression-free survival, weeks (range)	25.9 (2.3-162.7)	26.1 (3.7-162.7)	24.7 (2.3-103.4)	7.9 (4.6-11.1)
Prostate cancer-specific mortality	30 (48.4)	11 (31.4)	17 (68.0)	2 (100)
Median follow-up, weeks (range)	74.0 (4.7-182.1)	74.6 (4.7-182.1)	76.9 (21.4-144.9)	35.6 (16.4-54.7)

NOTE. Data are presented as No. (%) unless otherwise noted.

significantly associated with PSA response by logistic regression analysis (P = .119; Table 2 and Fig 2B), but was associated with a shorter median time to progression (16.1 weeks v 34.0 weeks; P = .013) and a shorter median survival (62.7 weeks v 144.9 weeks; P = .002; Figs 2C and 3B). Using proportional hazards regression modeling, PFS (HR, 2.07; 95% CI, 1.20 to 3.57; P = .009) and OS (HR, 3.26; 95% CI, 1.52 to 7.11; P = .002) were shorter in patients with *AR* CN gain; however, significance was lost upon inclusion of ctDNA burden in multivariable modeling (Figs 2C and 3B and Table 3).

Pathogenic *AR* LBD missense mutations were detected in cfDNA from 13% (eight of 62) of patients before therapy initiation and in an additional 15% (four of 26) of evaluable patients at disease progression (Figs 1C, 1D, and 2D and Data Supplement). Of the eight patients who had detectable *AR* LBD mutations before therapy, six did not have a PSA response, whereas two patients who harbored the

#### Torquato et al

TABLE 2.	Response to	Therapy:	Univariable and	Multivariable	Logistic	Regression	Analyses (N = 62	2)
----------	-------------	----------	-----------------	---------------	----------	------------	------------------	----

_		PSA Response (≥ 50%	6 decrease in PSA from baseline)		
		OR	95% CI	Р	
Response	Patients, No.		Univariable		
Prior abiraterone or enzalutamide	15	2.41	0.74 to 7.93	.146	
PSA ≥ 20 ng/mL	30	1.67	0.61 to 4.59	.323	
Age $\geq$ 72 years	32	1.32	0.48 to 3.63	.586	
Visceral metastasis	6	0.62	0.10 to 3.67	.598	
ctDNA high	27	3.17	1.11 to 9.05	.031	
AR CN gain and/or LBD mutation	34	1.80	0.65 to 5.01	.261	
AR LBD mutation	8	4.71	0.87 to 25.28	.072	
AR CN gain	32	2.27	0.81 to 6.34	.119	
TP53 mutation and/or CN loss	23	1.32	0.47 to 3.72	.602	
RB1 mutation and/or CN loss	17	2.35	0.75 to 7.35	.141	
<i>TP53</i> and <i>RB1</i> mutation and/or CN loss	6	7.73	0.85 to 70.65	.070	
PI3K pathway defect	15	8.53	2.09 to 34.81	.003	
WNT pathway defect	9	3.05	0.69 to 13.53	.143	
BRCA1/BRCA2/ATM mutation and/or CN loss	24	1.53	0.55 to 4.30	.417	
BRCA1/BRCA2/ATM truncating mutations	14	0.96	0.29 to 3.21	.953	
			Multivariable		
AR LBD mutation	8	4.88	0.85 to 28.08	.076	
TP53 and RB1 mutation and/or CN loss	6	5.40	0.56 to 52.15	.145	
PI3K pathway defect	15	7.09	1.40 to 35.94	.018	

NOTE. Significant *P* values are shown in bold. Multivariable analyses controlled for ctDNA high.

Abbreviations: AR, androgen receptor; ATM, ataxia-telangiectasia mutated gene; BRCA1/2, breast cancer gene 1/2; CN, copy number; ctDNA, circulating tumor DNA; LBD, ligand-binding domain; OR, odds ratio; PI3K, phosphoinositide 3-kinase; PSA, prostate-specific antigen; RB1, retinoblastoma-associated protein 1; TP53, tumor protein 53; WNT, wingless-type MMTV integration site.

H875Y mutation had PSA responses on abiraterone (Fig 2C). Using logistic regression analyses, *AR* LBD mutations were not significantly associated with a worse PSA response rate (P = .072; Table 2). However, pathogenic *AR* LBD missense mutations were associated with a worse 30% or more decline in PSA (OR, 6.00; 95% CI, 1.10 to 32.76; P = .039) that remained significant in multivariable logistic regression analyses (Data Supplement).

Median time to progression was shorter in patients who had a detectable *AR* LBD mutation than in patients without a detectable *AR* LBD mutation (11.4 weeks v 28.7 weeks; P = .021; Fig 2F). Using proportional hazards regression modeling, *AR* LBD mutations detected before therapy were associated with a shorter time to progression (HR, 2.39; 95% Cl, 1.11 to 5.14; P = .026), even when controlled for ctDNA burden (P = .020) and other variables (Table 3 and Data Supplement). However, detectable *AR* LBD mutations were not significantly associated with worse OS (P = .364; Table 3). *AR* CN gain and LBD mutations were not mutually exclusive in cfDNA (Fig 1C). Two *AR* mutations at different allelic frequencies—T878A at 9.4% and L702H at 1.5%—were detected in one patient who experienced disease progression on abiraterone plus prednisone who also had *AR* CN gain (Data Supplement). Studies support the idea that the *AR* L702H mutation mediates an acquired response to glucocorticoids, thereby providing rationale to switch from prednisone to dexamethasone.<sup>14,20,32</sup> In support of this notion, replacement of prednisone with dexamethasone resulted in a greater than 80% PSA decline for this patient (Data Supplement).

## TP53 and RB1

Genetic alterations in *TP53* are highly enriched in lethal prostate cancer<sup>8-11</sup> and have recently been shown to be associated with worse PFS and OS in patients treated with abiraterone and enzalutamide.<sup>17</sup> *TP53* was highly altered in patients' cfDNA (Figs 1C, 1D, and 3C); however, *TP53* 

			Pro	gression	-Free Su	rvival				<b>Overall</b>	Survival		
			Univariable			Multivariable			Univariable			Multivariable	
Variable	Patients, No.	H	95% CI	٩	H	95% CI	٩	HR	95% CI	٩	Ħ	95% CI	٩
Prior abiraterone or enzalutamide	15	1.17	0.63 to 2.14	.620	1.25	0.67 to 2.31	.480	1.51	0.71 to 3.24	.284	2.06	0.93 to 4.56	.074
PSA ≥ 20 ng/mL	30	1.43	0.83 to 2.45	.193	1.51	0.88 to 2.60	.137	1.44	0.70 to 2.95	.326	1.61	0.77 to 3.34	.204
Age ≥ 72 years	32	1.04	0.60 to 1.78	.895	1.07	0.62 to 1.84	.805	1.04	0.50 to 2.12	.925	1.11	0.54 to 2.28	.782
Visceral metastasis	9	1.29	0.51 to 3.26	.595	1.44	0.56 to 3.69	.450	2.50	0.95 to 6.63	.064	3.24	1.19 to 8.81	.021
ctDNA high	27	1.76	1.03 to 3.01	.039			1	2.92	1.40 to 6.11	.004			
AR CN gain and/or LBD mutation	34	2.00	1.15 to 3.47	.014	1.74	0.95 to 3.19	.072	2.95	1.36 to 6.36	900.	2.07	0.87 to 4.89	660.
AR LBD mutation	Ø	2.39	1.11 to 5.14	.026	2.51	1.15 to 5.45	.020	1.64	0.56 to 4.82	.364	1.69	0.57 to 4.97	.345
AR CN gain	32	2.07	1.20 to 3.57	600.	1.82	0.98 to 3.40	.060	3.26	1.52 to 7.00	.002	2.33	0.96 to 5.62	.060
TP53 mutation and/or CN loss	23	1.33	0.77 to 2.30	.314	1.26	0.73 to 2.19	.406	3.19	1.53 to 6.64	.002	2.70	1.27 to 5.72	600.
RB1 mutation and/or CN loss	17	0.94	0.52 to 1.70	.849	0.69	0.36 to 1.32	.265	1.47	0.68 to 3.18	.321	0.99	0.43 to 2.24	.974
TP53 and RB1 mutation and/or CN loss	9	1.66	0.71 to 3.91	.246	1.58	0.67 to 3.73	.299	4.50	1.79 to 11.28	.001	4.56	1.78 to 11.71	.002
PI3K pathway defect	15	1.77	0.97 to 3.22	.064	1.40	0.71 to 2.77	.327	3.64	1.69 to 7.86	.001	2.62	1.12 to 6.10	.026
WNT pathway defect	6	1.32	0.64 to 2.73	.450	1.13	0.54 to 2.39	.746	2.92	1.28 to 6.68	.011	2.36	0.995 to 5.60	.051
BRCA1/BRCA2/ATM mutation and/or CN loss	24	0.95	0.55 to 1.64	.855	0.89	0.52 to 1.54	.683	1.45	0.69 to 3.06	.326	1.32	0.62 to 2.85	.472
BRCA1/BRCA2/ATM truncating mutations	14	0.89	0.47 to 1.68	.715	0.86	0.45 to 1.63	.641	1.03	0.41 to 2.57	.948	0.97	0.39 to 2.44	.952
NOTE. Significant P values are shown in bold.	Multivariable ar	nalvses c	ontrolled for ct[	NA high									

**TABLE 3.** Progression-free Survival and Overall Survival: Univariable and Multivariable Cox Proportional Hazards Regression Analyses (N = 62) Prooression-Free Survival

Abbreviations: AR, androgen receptor; ATM, ataxia-telangiectasia mutated gene; BRCA1/2, breast cancer gene 1/2; CN, copy number; ctDNA, circulating tumor DNA; HR, hazard ratio; LBD, ligand binding domain; PI3K, phosphoinositide 3-kinase; PSA, prostate-specific antigen; RB1, retinoblastoma-associated protein 1; TP53, tumor protein 53; WNT, wingless-type MMTV integration site.

Downloaded from ascopubs.org by 31.40.211.25 on April 8, 2019 from 031.040.211.025 Copyright © 2019 American Society of Clinical Oncology. All rights reserved.



**FIG 1.** Genetic alterations detected in cell-free DNA (cfDNA) before therapy and best prostate-specific antigen (PSA) response. (A) Waterfall plot of best PSA response for all patients (N = 62) after therapy as determined by best percentage fold change in PSA. (B) Total number of protein-altering genetic changes in 46 genes detected by next-generation sequencing (NGS) of cfDNA from 62 patients before abiraterone (Abi) and enzalutamide (Enza) therapy. (C) Genetic alterations—copy number (CN) status, ClinVar pathogenic/likely pathogenic missense and germline mutations, and truncating mutations—in 46 genes detected by NGS of cfDNA from 62 patients before abiraterone (Abi) and enzalutamide (Enza) therapy. (C) Genetic alterations—copy number (CN) status, ClinVar pathogenic/likely pathogenic missense and germline mutations, and truncating mutations—in 46 genes detected by NGS of cfDNA from 62 patients before abiraterone and enzalutamide therapy in order of best PSA response. (D) Total number of genetic alterations—CN, ClinVar pathogenic/likely pathogenic missense and germline mutations, and truncating mutations—in 46 genes detected by NGS of cfDNA from 62 patients before abiraterone and enzalutamide therapy. AR, androgen receptor; ctDNA, circulating tumor DNA.

defects—pathogenic mutations and/or CN loss—were not associated with PSA response (P = .602) or PFS (P = .314; Tables 2 and 3). Conversely, median OS was shorter in patients with a *TP53* defect compared with patients without a detectable *TP53* defect (68.1 weeks v 134.9 weeks; P = .001; Fig 3D). Using proportional hazards regression modeling, *TP53* defects were associated with worse OS (HR, 3.19; 95% CI, 1.53 to 6.64; P = .002) that remained significant after adjusting for clinical variables (Table 3 and Data Supplement). Patients with both *TP53* and *RB1* defects had shorter median OS compared with patients with a *TP53* defect alone or with patients with intact *TP53* 





**FIG 2.** Progression-free survival (PFS): Pathogenic androgen receptor (*AR*) ligand-binding domain (LBD) mutations are associated with a shorter time to progression. (A) Kaplan-Meier method and log-rank test were used to determine median time to progression for patients who had high versus low circulating tumor DNA (ctDNA) before therapy. (B) Waterfall plot of best prostate-specific antigen (PSA) response for all patients (N = 62) after therapy as determined by best percentage fold change in PSA. *AR* copy number (CN) gain and *AR* LBD missense mutations were determined by deep next-generation sequencing (NGS) of cell-free DNA (cfDNA) before therapy. *AR* LBD missense mutations were included for patients with detectable mutations. Black arrow indicates patients with prior abiraterone or enzalutamide therapy.  $\chi^2$  analyses for a 30% or greater and 50% or greater PSA decrease. (C) Kaplan-Meier method and log-rank test were used to determine median time to progression for patients who had a gain in *AR* CN compared with patients who were *AR* CN neutral before therapy. The association of *AR* CN gain with PFS controlled for ctDNA burden using multivariable proportional hazards regression modeling. (C) Gene schematic illustrating pathogenic *AR* LBD mutations detected by targeted NGS of cfDNA before abiraterone and enzalutamide therapies. (E) Waterfall plot

(35.4 weeks v 77.4 weeks v 157.7 weeks; P < .001; Fig 3E). *TP53* defects in conjunction with *RB1* defects were associated with worse OS (HR, 4.50; 95% CI, 1.79 to 11.28; P = .001) that remained significant after adjusting for other variables (Table 3 and Data Supplement).

## PI3K and Wingless-Type MMTV Integration Site Pathways

PI3K pathway defects involving genetic alterations in PTEN-CN loss and/or truncating mutations-and PIK3-CA—CN gain and/or pathogenic missense mutation—were detected in nearly one quarter of patients before therapy (Figs 1C and 1D). Patients with PI3K pathway defects before therapy had a significantly shorter median survival (49.4 weeks v 134.9 weeks; P < .001) and worse OS (HR, 3.64; 95% CI, 1.69 to 7.86; P = .001), even after controlling for ctDNA burden (P = .026; Fig 3F, Table 3, and Data Supplement). PI3K pathway alterations were also associated with a worse PSA response that remained significant after adjusting for ctDNA burden (HR, 8.53; 95% CI, 2.09 to 34.81; P = .003; Table 2). Wingless-type MMTV integration site pathway defects involving genetic alterations in adenomatous polyposis coli-CN loss and/or truncating mutations—and β-catenin—CN gain and pathogenic missense mutations-were detected in nearly 15% of patients before to therapy (Figs 1C and 1D). Wingless-type MMTV integration site pathway defects were associated with a worse OS (HR, 2.92; 95% CI, 1.28 to 6.68; P = .011) using proportional hazards regression modeling; however, significance was lost after controlling for ctDNA burden (P = .051; Fig 3G and Table 3).

## BRCA1, BRCA2, and ATM

Men with lethal prostate cancer are more likely to have germline mutations in DNA repair genes<sup>33,34</sup>; however, the association of HDR gene defects with response to abiraterone and enzalutamide is conflicting.<sup>17,29,30</sup> Approximately one third of patients had germline and/or somatic deleterious mutations in or CN loss of *BRCA1*, *BRCA2*, or *ATM* before therapy, with some patients having more than one mutation (Figs 1C and 1D and Data Supplement). Collective ClinVar deleterious missense mutations, truncating mutations, and/or CN loss in *BRCA2*, *BRCA1*, or *ATM* were not significantly associated with PSA response (P = .417), PFS (P = .855), or OS (P = .326; Tables 2 and 3). Analysis of truncating mutations alone in *BRCA1*, *BRCA2*, and *ATM* did not increase prognostic significance.

## DISCUSSION

Liquid biopsies using cfDNA as a tumor analyte are rapidly being developed for cancer diagnostics of solid tumors.<sup>35-37</sup> When obtained concurrently, plasma-derived cfDNA is highly concordant with tissue biopsies for tumor-specific genetic alterations.<sup>38,39</sup> As a result of their advantages over traditional tissue biopsies, including the ease of accessibility for sequential monitoring of cancer dynamics and recapitulation of tumor heterogeneity, clinical development of cfDNA has the potential to advance prostate cancer precision medicine.<sup>40</sup>

Mechanisms of resistance to abiraterone and enzalutamide likely involve alterations to androgen-AR axis signaling. Previous studies have indicated that collective genetic aberrations to AR, including CN gain and mutations, are associated with worse outcomes in patients on abiraterone or enzalutamide therapies.<sup>14,15</sup> The value of AR LBD mutations alone as a predictive marker for response to enzalutamide and abiraterone in patients with mCRPC has yet to be fully established. A previous study demonstrated that patients with mCRPC harboring two or more AR mutations had worse outcomes on enzalutamide.<sup>20</sup> An additional retrospective study showed that AR mutations—L702H and T878A—were associated with shorter PFS and OS in postdocetaxel patients with mCRPC on abiraterone.<sup>19</sup> In contrast, a large prospective study reported that AR LBD mutations were not associated with time to progression on abiraterone or enzalutamide therapies in treatment-naïve patients with mCRPC.<sup>17</sup> In the current study, we found that AR LBD missense mutations detected in cfDNA before enzalutamide and abiraterone therapies were associated with a shorter PFS. but not PSA response or OS. Lack of a strong association with PSA response and OS lessens the likelihood that AR LBD mutations will be rigorous biomarkers for therapeutic decision making. Discrepancies between study findings may be a result of several factors, including prior therapies, study therapy, study design, specific AR LBD mutation, AR amplification, and disease burden. Prior therapies likely change the repertoire and incidence of AR LBD mutations.<sup>19,41</sup> As a result of their low individual prevalence, AR LBD mutations are often combined for analyses; however, studies support the idea that AR LBD mutations have distinct functional properties, including ligand promiscuity and agonistic activity, that mediate selective-therapy resistance.<sup>32,41</sup> Furthermore, the coincidence of other genetic alterations, including AR amplification or TP53 defects, and overall disease burden may be confounders. Future large-scale and multicenter prospective validation will be necessary to determine fully the roles of individual mutations in drug resistance.

AR CN gain as a single marker has been demonstrated to be associated with worse outcomes in patients with mCRPC

**FIG 2.** (Continued). of best PSA response for all patients (N = 62) after therapy as determined by best percentage fold change in PSA. *AR* LBD mutations were determined by deep NGS of cfDNA before therapy and listed below. Black arrow indicates patients with prior abiraterone or enzalutamide therapy.  $\chi^2$  analyses for a 30% or greater and 50% or greater PSA decrease. (F) Kaplan-Meier method and log-rank test were used to determine median time to progression for patients who were positive versus negative for *AR* LBD mutations before therapy. The association of *AR* LBD mutations with PFS controlled for ctDNA burden using multivariable proportional hazards regression modeling. DBD, DNA-binding domain; NTD, N-terminal domain.



JCO Precision Oncology

on abiraterone and enzalutamide.<sup>19,20</sup> A retrospective study reported that *AR* CN gain was associated with worse PFS and OS in men who were treated with enzalutamide or abiraterone for mCRPC.<sup>19</sup> Similarly, *AR* CN gain was also reported to be associated with a worse PSA response and PFS in patients on enzalutamide.<sup>20</sup> Our study also demonstrated an association of *AR* CN gain with PFS and OS; however, significance was lost in multivariable modeling, which is consistent with a previous report.<sup>17</sup> Clearly, additional prospective studies are needed to assess the clinical strength of *AR* CN gain as a predictive biomarker for therapeutic response to enzalutamide and abiraterone in patients with mCRPC.

In the current study, *TP53* and PI3K pathway defects were associated with worse OS. Deregulation of these pathways likely mediates resistance to androgen–AR axis therapies. Concurrent *TP53* and *RB1* defects are highly enriched in AR-independent neuroendocrine mCRPC compared with adenocarcinoma mCRPC.<sup>42</sup> Combined *TP53* and *RB1* loss has been shown to promote lineage switching from an AR-dependent to an AR-independent state<sup>41,43,44</sup> and consequent resistance to AR-targeted therapies. Similar to *TP53*, genetic alterations in *PTEN* are enriched in mCRPC compared with metastatic castration-sensitive prostate cancer and localized prostate cancer.<sup>11</sup>

Studies suggest that *PTEN* loss may mediate castration resistance by downregulating AR,<sup>25-28</sup> thereby supporting a rationale for combined inhibition of PI3K and AR- in *PTEN*-deficient mCRPCs.<sup>45,46</sup>

Association of pathogenic mutations in HDR genes with response to abiraterone and enzalutamide therapy is conflicting. A clinical trial in patients with mCRPC suggested that genetic alterations in HDR genes that were detected in metastatic biopsy tissue may be associated with longer PFS when on abiraterone therapy.<sup>29</sup> Concordant findings were observed in a second study that supported the idea that

patients with mCRPC harboring a germline *BRCA1/2* or *ATM* mutation may also have improved outcomes to abiraterone and enzalutamide.<sup>30</sup> In contrast, another study showed that truncating mutations in *BRCA2* and *ATM* detected in cfDNA were associated with a shorter time to progression on abiraterone and enzalutamide therapies in treatment-naïve patients with mCRPC.<sup>17</sup> In our study, collective somatic and germline genetic alterations were also not associated with worse outcomes to enzalutamide and abiraterone. Association differences may reflect variables, such as sample size, prior treatment status, disease burden, disease heterogeneity, somatic versus germline, and single versus dual loss. Certainly, additional prospective investigation is needed to determine the clinical significance of HDR mutations as predictive markers to abiraterone and enzalutamide therapies.

In the current study, many patients had detectable alterations that could serve as potential therapeutic targets. Previous studies have shown that patients with mCRPC with either germline or somatic mutations in HDR genes achieved significant responses to olaparib<sup>47</sup> and to abiraterone plus veliparib.<sup>29</sup> More than one quarter of patients in our study had a deleterious germline or somatic BRCA1, BRCA2, or ATM mutation detected before therapy or at disease progression, which suggests that these patients may benefit from therapies that target poly (ADP-ribose) polymerase or platinum-based chemotherapy.<sup>29,47,48</sup> In addition, immunotherapy trials have been largely unsuccessful in men with mCRPC<sup>49</sup>; however, rare responders have been reported.<sup>50</sup> A seminal clinical trial demonstrated that microsatellite instable cancers caused by mismatch repair (MMR) gene deficiency were sensitive to programmed death-1 blockade, perhaps because of the formation of neoantigens resulting from increased mutational burden.<sup>51</sup> Inactivation of MMR genes and elevated mutational burden have been detected in some men with aggressive prostate cancers.<sup>33,52,53</sup> One patient in our study had a detectable noncanonical MMR gene mutation in his cfDNA and a correspondingly high

FIG 3. Overall survival (OS): Tumor protein p53 (TP53) and phosphoinositide 3-kinase (PI3K) pathway defects are associated with worse OS. (A) Kaplan-Meier method and log-rank test were used to determine median OS for patients who had high versus low circulating tumor DNA (ctDNA) before therapy. (B) Kaplan-Meier method and log-rank test were used to determine median OS for patients who had androgen receptor (AR) copy number (CN) gain before therapy. Association of AR CN gain with OS controlled for ctDNA burden using multivariable proportional hazards regression modeling. (C) Gene schematic illustrating deleterious TP53 mutations detected by deep next-generation sequencing (NGS) of cellfree DNA (cfDNA) before abiraterone and enzalutamide therapies and at disease progression while on therapy. (D) Kaplan-Meier method and log-rank test were used to determine median OS for patients who had TP53 defects—CN loss and or ClinVar pathogenic/likely pathogenic mutations—before therapy. Association of TP53 defects with OS controlled for ctDNA burden using multivariable proportional hazards regression modeling. (E) Kaplan-Meier method and log-rank test were used to determine median OS for patients who had both TP53 and retinoblastomaassociated protein 1 (RB1) defects compared with patients who had TP53 defects but were RB1 intact—CN loss and or ClinVar pathogenic/likely pathogenic mutations—before therapy. Association of dual TP53 and RB1 defects with OS controlled for ctDNA burden using multivariable proportional hazards regression modeling. (F) Kaplan-Meier method and log-rank test were used to determine median OS for patients who had PI3K pathway defects—CN loss and/or truncating mutations in phosphatase and tensin homolog and/or CN gain of PIK3CA—before therapy. Association of PI3K defects with OS controlled for ctDNA burden using multivariable proportional hazards regression modeling. (G) Kaplan-Meier method and log-rank test were used to determine median OS for patients who had wingless-type MMTV integration site (WNT) pathway defects-CN loss and/or truncating mutations in adenomatous polyposis coli and/or CN gain and/or pathogenic missense mutations in β-catenin—before therapy. Association of WNT defects with OS controlled for ctDNA burden using multivariable proportional hazards regression modeling. P53, P53 DNA-binding domain; P53T, P53 transactivation motif; P53Tetra, P53 tetramerisation motif.

mutational burden that suggested that he may be an ideal candidate for checkpoint immunotherapy. This study supports the idea that cfDNA may be a useful analyte for directing clinical decisions in prostate cancer precision medicine.

Several limitations to our study exist. Of note, the small sample size precluded multivariable analyses that incorporated more than two variables and analyses by therapy subgroup. Consistent with other reports, patients with prior exposure to abiraterone and enzalutamide experienced worse outcomes.<sup>54-59</sup> Statistical significance was not reached, likely because of the small overall sample size and the few patients with prior therapy. In addition, the small size precluded any definitive conclusions pertaining to the association of AR LBD mutations with outcomes. Larger prospective studies will be needed to validate our findings. Samples were obtained from two hospitals, and future prospective studies would benefit from the inclusion of a larger number of institutions. Future prospective studies would also be strengthened by radiologic confirmation of progression for every patient. An additional limitation was the variability of cfDNA input for NGS among patients. NGS protocols were adjusted on the basis of total input, but for patients with low input the lack of genetic alteration detection was considered indeterminate as opposed to negative. In addition, mutations in such genes as

#### **AFFILIATIONS**

<sup>1</sup>Johns Hopkins School of Medicine, Baltimore, MD <sup>2</sup>Johns Hopkins University, Baltimore, MD

#### **CORRESPONDING AUTHOR**

Paula J. Hurley, PhD, The James Buchanan Brady Urological Institute, Department of Urology, Johns Hopkins University, CRBII 146, 1550 Orleans Ave, Baltimore, MD 21231; Twitter: @HurleyLab; @brady\_ urology; @HopkinsMedicine; e-mail: phurley2@jhmi.edu.

### **EQUAL CONTRIBUTION**

S.T. and A.P. contributed equally to this work.

#### SUPPORT

Supported by the Department of Defense CDMRP Prostate Cancer Research Program PC130991 Grant No. W81XWH-14-1-0284 (P.J.H., B.H.P., B.J.T., A.G.), the Patrick C. Walsh Foundation (B.H.P., P.J.H.), the Prostate Cancer Foundation Young Investigator Award (E.S.A.), the Department of Defense Prostate Cancer Research Program Grant No. W81XWH-15-2-0050 (E.S.A.), National Cancer Institute Grants No. R01-CA194024 and R01-CA214494 (B.H.P.), the Breast Cancer Research Foundation (B.H.P.), and the Commonwealth Foundation (B.H.P.). B.H.P. received support from Susan G. Komen as a Komen Scholar.

#### AUTHOR CONTRIBUTIONS

Conception and design: Samantha Torquato, Emmanuel S. Antonarakis, Ben H. Park, Paula J. Hurley Financial support: Paula J. Hurley Administrative support: Paula J. Hurley *TP53* and *ATM* detected in cfDNA at low allelic frequencies may be false positives as a result of clonal hematopoiesis.<sup>60</sup> Corresponding tissue was not available for all samples to confirm *TP53* status, and future studies will examine both prostate tumor tissue and blood leukocytes for genetic alterations. A final limitation was our inability to evaluate *AR* splice variants, including *AR*-V7, because of the requirement of circulating tumor cells or whole-blood RNA. The presence of AR-V7 is certainly another established mechanism of primary and acquired resistance to next-generation hormonal therapies.<sup>21-23</sup> Future studies should aim to simultaneously analyze the full complement of *AR* aberrations, including gene mutations, amplifications, genomic structural rearrangements, and mRNA splice variants, from a single liquid biopsy.

In summary, our findings indicate ctDNA burden was highly associated with worse outcomes to enzalutamide and abiraterone. Association of *AR* status with outcomes was not robust and will need additional prospective validation. *TP53* loss, especially in the context of concurrent *RB1* defects, and PI3K pathway defects were associated with worse OS. These studies provide the rationale for larger prospective multi-institutional studies to additionally assess the clinical utility of integrating genetic alterations detected in cfDNA for the optimal management of metastatic prostate cancer.

Provision of study materials or patients: Patricia Valda Toro, Michael A. Carducci, Channing J. Paller, Samuel R. Denmeade, Bruce Kressel, Mario A. Eisenberger, Emmanuel S. Antonarakis, Paula J. Hurley
Collection and assembly of data: Samantha Torquato, Alexa Goldstein, Patricia Valda Toro, John L. Silberstein, Justin Lee, Mary Nakazawa, Ian Waters, Daniel Shinn, Taylor Groginski, Brian W. Simons, Hamda Khan, Channing J. Paller, Samuel R. Denmeade, Bruce Kressel, Mario A. Eisenberger, Emmanuel S. Antonarakis, Paula J. Hurley
Data analysis and interpretation: Samantha Torquato, Aparna Pallavajjala, David Chu, Robert M. Hughes, Hamda Khan, Zhaoyong Feng, Michael A. Carducci, Channing J. Paller, Emmanuel S. Antonarakis, Bruce J. Trock, Ben H. Park, Paula J. Hurley
Manuscript writing: All authors
Final approval of manuscript: All authors

Accountable for all aspects of the work: All authors

AUTHORS' DISCLOSURES OF POTENTIAL CONFLICTS OF INTEREST The following represents disclosure information provided by authors of this manuscript. All relationships are considered compensated. Relationships are self-held unless noted. I = Immediate Family Member, Inst = My Institution. Relationships may not relate to the subject matter of this manuscript. For more information about ASCO's conflict of interest policy, please refer to www.asco.org/rwc or po.ascopubs.org/site/ifc.

#### Michael A. Carducci

**Consulting or Advisory Role:** Astellas Pharma, AbbVie, Genentech, Pfizer, Foundation Medicine

**Research Funding:** Bristol-Myers Squibb (Inst), Pfizer (Inst), AstraZeneca (Inst), Gilead Sciences (Inst), EMD Serono (Inst)

Channing J. Paller Consulting or Advisory Role: Dendreon Research Funding: Eli Lilly (Inst)

Samuel R. Denmeade Stock and Other Ownership Interests: Sophiris Bio Consulting or Advisory Role: Sophiris Bio Travel, Accommodations, Expenses: Sophiris Bio

Mario A. Eisenberger Leadership: Veru Stock and Other Ownership Interests: Veru

Honoraria: Sanofi, Pfizer

Consulting or Advisory Role: Astellas Pharma, Ipsen, Bayer, Sanofi, Pfizer Research Funding: Sanofi, Tokai Pharmaceuticals, Genentech Travel, Accommodations, Expenses: Bayer, Astellas Pharma, Sanofi, Pfizer, Veru

#### **Emmanuel S. Antonarakis**

Honoraria: Sanofi, Dendreon, Medivation, Janssen Biotech, ESSA Pharma, Astellas Pharma, Merck, AstraZeneca, Clovis Oncology Consulting or Advisory Role: Sanofi, Dendreon, Medivation, Janssen Biotech, ESSA Pharma, Astellas Pharma, Merck, AstraZeneca, Clovis Oncology

**Research Funding:** Janssen Biotech (Inst), Johnson & Johnson (Inst), Sanofi (Inst), Dendreon (Inst), Aragon Pharmaceuticals (Inst), Exelixis (Inst), Millennium Pharmaceuticals (Inst), Genentech (Inst), Novartis (Inst), Astellas Pharma (Inst), Tokai Pharmaceuticals (Inst), Merck (Inst), AstraZeneca (Inst), Clovis Oncology (Inst), Constellation Pharmaceuticals (Inst)

Patents, Royalties, Other Intellectual Property: Coinventor of a biomarker technology licensed to Qiagen

Travel, Accommodations, Expenses: Sanofi, Dendreon, Medivation

Bruce J. Trock Consulting or Advisory Role: GenomeDx, Myriad Genetics Research Funding: Myriad Genetics, MDxHealth

#### Ben H. Park

Leadership: Loxo Pharmaceuticals

Stock and Other Ownership Interests: Loxo Pharmaceuticals Consulting or Advisory Role: Horizon Discovery, Foundation Medicine, Loxo Pharmaceuticals, Casdin Capital, H3 Biomedicine, Jackson Laboratory for Genomic Medicine, Eli Lilly Speakers' Bureau: AstraZeneca

Research Funding: Foundation Medicine, AbbVie, Pfizer Patents, Royalties, Other Intellectual Property: Royalties paid through inventions at Johns Hopkins University by Horizon Discovery Travel, Accommodations, Expenses: Eli Lilly, Loxo Pharmaceuticals

#### Paula J. Hurley

Stock and Other Ownership Interests: Loxo Pharmaceuticals (I)
Honoraria: Foundation Medicine (I), Roche (I), H3 Biomedicine (I), Casdin Capital (I), Loxo Pharmaceuticals (I), Eli Lilly (I)
Consulting or Advisory Role: Loxo Pharmaceuticals (I), Foundation
Medicine (I), Roche (I), H3 Biomedicine (I), Casdin Capital (I), Lilly (I)
Patents, Royalties, Other Intellectual Property: Cell lines at Horizon (I)
No other potential conflicts of interest were reported.

#### ACKNOWLEDGMENT

We thank the patients who consented to participate in this study. We thank Ashley E. Ross and Edward M. Schaeffer for helpful discussions. We also thank Jennifer Meyers and Michael Rongione at the Johns Hopkins next-generation sequencing core.

#### REFERENCES

- 1. de Bono JS, Logothetis CJ, Molina A, et al: Abiraterone and increased survival in metastatic prostate cancer. N Engl J Med 364:1995-2005, 2011
- 2. Ryan CJ, Smith MR, de Bono JS, et al: Abiraterone in metastatic prostate cancer without previous chemotherapy. N Engl J Med 368:138-148, 2013
- 3. Scher HI, Fizazi K, Saad F, et al: Increased survival with enzalutamide in prostate cancer after chemotherapy. N Engl J Med 367:1187-1197, 2012
- 4. Beer TM, Armstrong AJ, Rathkopf DE, et al: Enzalutamide in metastatic prostate cancer before chemotherapy. N Engl J Med 371:424-433, 2014
- 5. Cancer Genome Atlas Research Network: Comprehensive molecular characterization of urothelial bladder carcinoma. Nature 507:315-322, 2014
- 6. Baca SC, Prandi D, Lawrence MS, et al: Punctuated evolution of prostate cancer genomes. Cell 153:666-677, 2013
- 7. Taylor BS, Schultz N, Hieronymus H, et al: Integrative genomic profiling of human prostate cancer. Cancer Cell 18:11-22, 2010
- 8. Grasso CS, Wu YM, Robinson DR, et al: The mutational landscape of lethal castration-resistant prostate cancer. Nature 487:239-243, 2012
- 9. Kumar A, Coleman I, Morrissey C, et al: Substantial interindividual and limited intraindividual genomic diversity among tumors from men with metastatic prostate cancer. Nat Med 22:369-378, 2016
- 10. Robinson D, Van Allen EM, Wu YM, et al: Integrative clinical genomics of advanced prostate cancer. Cell 161:1215-1228, 2015 [Erratum: Cell 162:454, 2015]
- 11. Abida W, Armenia J, Gopalan A, et al: Prospective genomic profiling of prostate cancer across disease states reveals germline and somatic alterations that may affect clinical decision making. JCO Precis Oncol doi:10.1200/P0.17.00029
- 12. Cerami E, Gao J, Dogrusoz U, et al: The cBio cancer genomics portal: An open platform for exploring multidimensional cancer genomics data. Cancer Discov 2:401-404, 2012
- 13. Gao J, Aksoy BA, Dogrusoz U, et al: Integrative analysis of complex cancer genomics and clinical profiles using the cBioPortal. Sci Signal 6:pl1, 2013
- 14. Romanel A, Gasi Tandefelt D, Conteduca V, et al: Plasma AR and abiraterone-resistant prostate cancer. Sci Transl Med 7:312re10, 2015
- 15. Azad AA, Volik SV, Wyatt AW, et al: Androgen receptor gene aberrations in circulating cell-free DNA: Biomarkers of therapeutic resistance in castration-resistant prostate cancer. Clin Cancer Res 21:2315-2324, 2015
- 16. Goldstein A, Toro PV, Lee J, et al: Detection fidelity of AR mutations in plasma derived cell-free DNA. Oncotarget 8:15651-15662, 2017
- 17. Annala M, Vandekerkhove G, Khalaf D, et al: Circulating tumor DNA genomics correlate with resistance to abiraterone and enzalutamide in prostate cancer. Cancer Discov 8:444-457, 2018
- Salvi S, Casadio V, Conteduca V, et al: Circulating cell-free AR and CYP17A1 copy number variations may associate with outcome of metastatic castrationresistant prostate cancer patients treated with abiraterone. Br J Cancer 112:1717-1724, 2015
- Conteduca V, Wetterskog D, Sharabiani MTA, et al: Androgen receptor gene status in plasma DNA associates with worse outcome on enzalutamide or abiraterone for castration-resistant prostate cancer: A multi-institution correlative biomarker study. Ann Oncol 28:1508-1516, 2017

- 20. Wyatt AW, Azad AA, Volik SV, et al: Genomic alterations in cell-free DNA and enzalutamide resistance in castration-resistant prostate cancer. JAMA Oncol 2:1598-1606, 2016
- 21. Antonarakis ES, Lu C, Wang H, et al: AR-V7 and resistance to enzalutamide and abiraterone in prostate cancer. N Engl J Med 371:1028-1038, 2014
- 22. Qu F, Xie W, Nakabayashi M, et al: Association of AR-V7 and prostate-specific antigen RNA levels in blood with efficacy of abiraterone acetate and enzalutamide treatment in men with prostate cancer. Clin Cancer Res 23:726-734, 2017
- Antonarakis ES, Lu C, Luber B, et al: Clinical significance of androgen receptor splice variant-7 mRNA detection in circulating tumor cells of men with metastatic castration-resistant prostate cancer treated with first- and second-line abiraterone and enzalutamide. J Clin Oncol 35:2149-2156, 2017
- 24. Henzler C, Li Y, Yang R, et al: Truncation and constitutive activation of the androgen receptor by diverse genomic rearrangements in prostate cancer. Nat Commun 7:13668, 2016
- 25. Shen MM, Abate-Shen C: Pten inactivation and the emergence of androgen-independent prostate cancer. Cancer Res 67:6535-6538, 2007
- 26. Jiao J, Wang S, Qiao R, et al: Murine cell lines derived from Pten null prostate cancer show the critical role of PTEN in hormone refractory prostate cancer development. Cancer Res 67:6083-6091, 2007
- 27. Mulholland DJ, Tran LM, Li Y, et al: Cell autonomous role of PTEN in regulating castration-resistant prostate cancer growth. Cancer Cell 19:792-804, 2011
- Carver BS, Chapinski C, Wongvipat J, et al: Reciprocal feedback regulation of PI3K and androgen receptor signaling in PTEN-deficient prostate cancer. Cancer Cell 19:575-586, 2011
- 29. Hussain M, Daignault-Newton S, Twardowski PW, et al: Targeting androgen receptor and DNA repair in metastatic castration-resistant prostate cancer: Results from NCI 9012. J Clin Oncol 36:991-999, 2018
- Antonarakis ES, Changxue L, Luber B, et al: Germline DNA-repair gene mutations and outcomes in men with metastatic castration-resistant prostate cancer receiving first-line abiraterone and enzalutamide. Eur Urol 74:218-225, 2018
- Salvi S, Casadio V, Conteduca V, et al: Circulating AR copy number and outcome to enzalutamide in docetaxel-treated metastatic castration-resistant prostate cancer. Oncotarget 7:37839-37845, 2016
- Lallous N, Volik SV, Awrey S, et al: Functional analysis of androgen receptor mutations that confer anti-androgen resistance identified in circulating cell-free DNA from prostate cancer patients. Genome Biol 17:10, 2016
- 33. Pritchard CC, Mateo J, Walsh MF, et al: Inherited DNA-repair gene mutations in men with metastatic prostate cancer. N Engl J Med 375:443-453, 2016
- Na R, Zheng SL, Han M, et al: Germline mutations in ATM and BRCA1/2 distinguish risk for lethal and indolent prostate cancer and are associated with early age at death. Eur Urol 71:740-747, 2017
- Cohen JD, Li L, Wang Y, et al: Detection and localization of surgically resectable cancers with a multi-analyte blood test. Science 359:926-930, 2018
- 36. Leary RJ, Sausen M, Kinde I, et al: Detection of chromosomal alterations in the circulation of cancer patients with whole-genome sequencing. Sci Transl Med 4:162ra154, 2012
- 37. Webb S: The cancer bloodhounds. Nat Biotechnol 34:1090-1094, 2016 [Erratum: Nat Biotechnol 35:178, 2017]
- Wyatt AW, Annala M, Aggarwal R, et al: Concordance of circulating tumor DNA and matched metastatic tissue biopsy in prostate cancer. J Natl Cancer Inst 110:78-86, 2018
- Beaver JA, Jelovac D, Balukrishna S, et al: Detection of cancer DNA in plasma of patients with early-stage breast cancer. Clin Cancer Res 20:2643-2650, 2014
- 40. Schweizer MT, Antonarakis ES: Liquid biopsy: Clues on prostate cancer drug resistance. Sci Transl Med 7:312fs45, 2015
- 41. Watson PA, Arora VK, Sawyers CL: Emerging mechanisms of resistance to androgen receptor inhibitors in prostate cancer. Nat Rev Cancer 15:701-711, 2015
- 42. Beltran H, Prandi D, Mosquera JM, et al: Divergent clonal evolution of castration-resistant neuroendocrine prostate cancer. Nat Med 22:298-305, 2016
- Ku SY, Rosario S, Wang Y, et al: Rb1 and Trp53 cooperate to suppress prostate cancer lineage plasticity, metastasis, and antiandrogen resistance. Science 355:78-83, 2017
- Mu P, Zhang Z, Benelli M, et al: SOX2 promotes lineage plasticity and antiandrogen resistance in TP53- and RB1-deficient prostate cancer. Science 355:84-88, 2017
- 45. Jamaspishvili T, Berman DM, Ross AE, et al: Clinical implications of PTEN loss in prostate cancer. Nat Rev Urol 15:222-234, 2018
- 46. de Bono JS, De Giorgi U., Massard C, et al: PTEN loss as a predictive biomarker for the Akt inhibitor ipatasertib combined with abiraterone acetate in patients with metastatic castration-resistant prostate cancer (mCRPC). Ann Oncol 27:7180, 2016 (suppl 6)
- 47. Mateo J, Carreira S, Sandhu S, et al: DNA-repair defects and olaparib in metastatic prostate cancer. N Engl J Med 373:1697-1708, 2015
- 48. Cheng HH, Pritchard CC, Boyd T, et al: Biallelic inactivation of BRCA2 in platinum-sensitive metastatic castration-resistant prostate cancer. Eur Urol 69:992-995, 2016
- 49. Brahmer JR, Drake CG, Wollner I, et al: Phase I study of single-agent anti-programmed death-1 (MDX-1106) in refractory solid tumors: Safety, clinical activity, pharmacodynamics, and immunologic correlates. J Clin Oncol 28:3167-3175, 2010
- Kwon ED, Drake CG, Scher HI, et al: Ipilimumab versus placebo after radiotherapy in patients with metastatic castration-resistant prostate cancer that had progressed after docetaxel chemotherapy (CA184-043): A multicentre, randomised, double-blind, phase 3 trial. Lancet Oncol 15:700-712, 2014
- 51. Le DT, Uram JN, Wang H, et al: PD-1 blockade in tumors with mismatch-repair deficiency. N Engl J Med 372:2509-2520, 2015
- 52. Guedes LB, Antonarakis ES, Schweizer MT, et al: MSH2 loss in primary prostate cancer. Clin Cancer Res 23:6863-6874, 2017
- 53. Pritchard CC, Morrissey C, Kumar A, et al: Complex MSH2 and MSH6 mutations in hypermutated microsatellite unstable advanced prostate cancer. Nat Commun 5:4988, 2014
- 54. Loriot Y, Bianchini D, Ileana E, et al: Antitumour activity of abiraterone acetate against metastatic castration-resistant prostate cancer progressing after docetaxel and enzalutamide (MDV3100). Ann Oncol 24:1807-1812, 2013
- Noonan KL, North S, Bitting RL, et al: Clinical activity of abiraterone acetate in patients with metastatic castration-resistant prostate cancer progressing after enzalutamide. Ann Oncol 24:1802-1807, 2013
- Azad AA, Eigl BJ, Murray RN, et al: Efficacy of enzalutamide following abiraterone acetate in chemotherapy-naive metastatic castration-resistant prostate cancer patients. Eur Urol 67:23-29, 2015
- 57. Brasso K, Thomsen FB, Schrader AJ, et al: Enzalutamide antitumour activity against metastatic castration-resistant prostate cancer previously treated with docetaxel and abiraterone: A multicentre analysis. Eur Urol 68:317-324, 2015

#### Torquato et al

- Maughan BL, Luber B, Nadal R, et al: Comparing sequencing of abiraterone and enzalutamide in men with metastatic castration-resistant prostate cancer: A retrospective study. Prostate 77:33-40, 2017
- Emamekhoo H, Barata PC, Edwin NC, et al: Evaluation of response to enzalutamide consecutively after abiraterone acetate/prednisone failure in patients with metastatic castration-resistant prostate cancer. Clin Genitourin Cancer 16:429-436, 2018

----

60. Hu Y, Ulrich BC, Supplee J, et al: False positive plasma genotyping due to clonal hematopoiesis. Clin Cancer Res 24:4437-4443, 2018